

Formulation, Optimization and Evaluation of *in-situ* Gelling Liquid Oral Formulation of a Novel Antidiabetic Drug: Canagliflozin

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ABSTRACT

Aim: The present research work involves the development and optimization of an *in-situ* gel forming formulation by face centered central composite response surface design methodology to increase buoyancy and controlled release of a novel antidiabetic drug; Canagliflozin hemihydrate, having absorption window in upper part of gastrointestinal tract. **Materials and Methods:** The drug and excipient compatibility studies were performed by subjecting to Fourier Transform Infra-red spectroscopy and Differential Scanning Calorimetry analysis. Nine primary formulations suggested by Design Expert software were formulated and subjected for the evaluation by measurement of rheological properties, buoyancy studies, drug content and *in-vitro* drug release studies. The optimization method adopted in the present study was face centered central composite response surface design. **Result:** The effect of two independent factors namely concentration of sodium alginate and concentration of gas generating agent calcium carbonate on responses like floating lag time, viscosity and *in-vitro* drug release were considered for the optimization by plotting three dimensional surface response plots. The results of these studies were subjected to analysis by software to obtain an optimized formulation. The software suggested the optimized formulation must consists of 0.868 % of sodium alginate and 0.957 % of calcium carbonate and these compositions are much near to the composition of SF5. **Conclusion:** By considering the effect of calcium cations on the gelling properties of various concentration of sodium alginate, it was concluded that the effective gelling was achieved in the formulation comprised of 1 % sodium alginate with 0.5 %, 1.0 % and 1.5 % of calcium carbonate.

Key words: Gastric retention, Antidiabetic, Canagliflozin Hemihydrate, Sodium Alginate, Optimization.

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INTRODUCTION

Short Gastric Residence Time (GRT) and unpredictable Gastric Emptying Time (GET) are the physiological adversities challenge the development of rate controlled oral drug delivery systems. To overcome these, certain scientific and technological advancements have been made.¹ It is always better to develop ideal drug delivery systems that will be a single dose for the duration of treatment and deliver the drug directly at the site of action.

The bioavailability of drugs with narrow absorption window in stomach (like captopril, metformin, riboflavin etc.) can be

increased thus reducing the dose. The stomach with a highly acidic pH is not free from microbial colonization like of *Helicobacter pylori*^{2,3} (leads to ulcer) or other organisms. By targeting drugs like amoxicillin, metronidazole,³ tetracycline³ and anti-histamines help in eradication of pathogenic organisms and avoid dose dumping.

Gastro-retentive Drug Delivery System (GRDDS) has gained immense popularity in the field of oral drug delivery recently. It is a widely employed approach to retain the dosage form in the stomach for an extended period of time and release the



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drug slowly that can address many challenges associated with conventional oral delivery, including poor bioavailability. Different innovative approaches like magnetic field assisted gastro-retention, plug type swelling system, muco-adhesion technique, floating system with or without effervescence are being applied to fabricate GRDDS.

Conventional oral dosage forms pose low bioavailability problems due to their rapid gastric transition from stomach. Drugs which produce their local action in stomach get rapidly emptied and do not get enough residence time in stomach. So, frequency of dose administration in such cases is increased. To avoid this problem, various efforts have been made to prolong the retention time of drug delivery system.

Several approaches are currently utilized in the prolongation of the GRT, including Floating Drug Delivery Systems (FDDS), are also known as Hydro Dynamically Balanced Systems (HBS), swelling and expanding systems, polymeric bio-adhesive systems,⁴ modified shape systems and high-density systems. The various buoyant preparations include hollow microspheres^{5,6} (micro balloons), granules, powders, capsules, tablets and laminated films.

Sodium alginate is widely used in pharmaceutical formulations in beads, matrix tablets and microcapsules. In presence of di- and trivalent metal ions, dilute solutions of sodium alginate undergoes gelation by a process involving consecutive guluronic residues in the guluronic acid (G) blocks of the alginate chain. This property is successfully utilized by many researchers for the formulation of novel drug delivery systems.⁷

Canagliflozin hemihydrate is the recently FDA approved first oral agent in a novel class of diabetes drugs known as sodium–glucose co-transporter-2 (SGLT-2) inhibitors. Canagliflozin is rapidly absorbed in the gastrointestinal (GI) tract. It has a relative oral bioavailability of 65%. It can be taken without regard to food, but it is recommended that it be taken before the first meal of the day.⁸ Solubility of Canagliflozin is of major concern as it is insoluble in water. With all these properties of the Canagliflozin, in this work we have tried to develop a dosage form which can address and overcome the challenges in achieving efficient drug delivery.

MATERIALS AND METHODS

Materials

Canagliflozin was purchased from Watson Pharma Private Limited, Mumbai, India, Sodium alginate was a gift sample from Anthem Bio Sciences Private Limited, Bengaluru, India and all other chemicals and reagents used were of research grade.

Methods

Drug identification and authentication

Canagliflozin identified and authenticated by determining melting point (Lab India, Visual Melting Range Apparatus, MR-VIS[®]), solubility, Fourier-Transform Infrared spectroscopy (FTIR) (Shimadzu IR Affinity-1S instrument) and Differential Scanning Calorimetry (DSC) (Mettler star SW-9.01) spectral analysis.

Drug polymer compatibility studies

Polymer and drug Canagliflozin compatibility study was carried out by subjecting the drug and polymer (sodium alginate) individually as well as in physical mixture into FTIR and DSC.

Fourier transform infrared spectroscopy

FTIR studies were carried out in Shimadzu IR Affinity-1S instrument. The individual polymer and pure drug were placed on the germanium prism and FTIR spectra were recorded in the scanning measurement range of 600 to 4000 cm^{-1} . In the same manner FTIR spectra were recorded for the physical mixtures of polymer and drug (1:1 ratio).

Differential scanning calorimetry

DSC studies were carried out in Mettler star SW-9.01 instrument. About 1-3 mg of polymer, pure drug and the physical mixtures of polymer and drug (1:1 ratio) were placed in aluminum pans and the pans were closed by crimping the lids. The filled aluminum pans and empty sealed pan were placed in a sample holder where empty pan acts as a reference. The thermal behavior of samples was investigated under nitrogen purging at scanning rate of 10°C/m, covering the temperature range of 30-360°C. Heat runs for each sample were set from 30-360°C using nitrogen as purging gas and the samples were analysed.

Optimization of floating *in-situ* gelling formulation using Design Expert software

A two-level three-response surface face centered central composite design for experimentation is utilized in present study. Statistical experimental design was performed using DESIGN EXPERT[®] version 11.0 (Stat-Ease Inc., Minneapolis, USA) software. Response surface graphs were used to show the factor interaction between the considered variables. Selected independent variables studied were the concentration of sodium alginate (X1); and Calcium carbonate (X2) added to the formulation. Three factorial levels coded for low, medium and high settings (-1, 0 and +1, respectively) were considered for two independent variables. The selected dependent variables investigated were the viscosity (Y1), floating lag time (Y2),

percentage of drug released at 1st h (Y3) and 8th h (Y4). A total of 9 (Table 4) experimental runs were required for analyzing the interaction of each level on formulation characters and to optimize. Results obtained for different evaluation parameters were subjected to software for analysis and further optimization of the formulations.⁹⁻¹²

Formulation of gastro retentive *in-situ* gelling liquid

Sodium alginate, at solution concentrations of 0.5–1.5% (w/v) were prepared in de-ionized water by heating to 60°C with continuous stirring. After cooling below 40°C, various concentrations of calcium carbonate and drug solutions were added and dispersed well with continuous stirring. The resulting sodium alginate *in-situ* gel solution containing Canagliflozin was finally stored in amber color narrow mouth bottles until further use.¹³ (Table 1)

Evaluation

Rheological properties of the formulations

The viscosities of formulations were determined by Brookfield viscometer DV-III using spindle no. 21. Viscosity was measured at different Rotations Per Min (RPM) to get corresponding angular velocities and the findings are depicted in Table 2.¹³

Drug content

One ml of the formulation added to 10 ml volumetric flask with 7.5 ml of methanol and stirred for 1 h on magnetic stirrer and made up to the mark with methanol. The solution was filtered and 1 ml of this solution is transferred to 10 ml volumetric flask and made up the mark with mobile phase. The amount of Canagliflozin was estimated by RP-HPLC method. Estimation of Canagliflozin was carried out using Shimadzu, Prominence LC system comprising COSMIC-SIL 100 C₁₈ 5 µm column at room temperature with mobile phase containing HPLC grade Acetonitrile and Water in the ratio 70:30, pH adjusted to 3.0 using *ortho* phosphoric acid. The flow rate of the mobile phase was 1.0 ml/m, injection volume 20 µl and detection wavelength 282 nm. Each determination carried out in triplicate and results are given in Table 2.¹³

Buoyancy studies

The time taken by the *in-situ* gel to buoyant on the surface of the medium (floating lag time) was noted. The time duration the formulation constantly floated on the surface of the dissolution medium (duration of floating) was noted.¹³ (Table 2)

In-vitro drug release

The cumulative *in-vitro* drug release (CDR) was determined using USP dissolution apparatus II having 900 ml of 0.1N Hydrochloric acid of pH 1.2 (0.1N HCl

of pH 1.2) with 0.75% w/v of sodium lauryl sulphate. The medium temperature was maintained at 37°C. Ten ml of *in-situ* gelling formulation was placed into the petridish of 4.5 cm internal diameter and petridish containing formulation was kept in the dissolution vessel. Then carefully transferred the dissolution medium into the dissolution vessel without disturbing the formulation. At each time interval, a precisely measured sample of the dissolution medium was removed and replenished with prewarmed (37°C) fresh medium. Amount of Canagliflozin in withdrawn samples was estimated by RP-HPLC method as mentioned in drug content determination. Interference from the excipients was negligible. Each study was conducted in triplicate till 8 h.¹³ (Table 3)

Analysis of drug release mechanism

Several kinetics models describe drug dissolution from immediate and modified release dosage forms. There are several models to represent the drug dissolution profiles where *f* is a function of *t* (time) related to the amount of drug dissolved from the pharmaceutical dosage system. The quantitative interpretation of the values obtained in the dissolution assay is facilitated by the usage of a generic equation that mathematically translates the dissolution curve in function of some parameters related with the pharmaceutical dosage forms. All the formulations were analysed for release kinetics by different models Zero order, First order, Higuchi's diffusion Hixson Crowell and Korsmeyer-Peppas models.^{14,15} (Table 4)

Scanning electron microscopy

Scanning Electron Microscopic evaluation was carried out using TESCAN VEGA 3 instrument. The sample housing is cleaned with a damp wipe and a SEM substrate (stub) is prepared for receiving a filter by applying a double-sided, sticky conductive carbon pad to the stub surface. The underside of the stub is labeled with the sample number using a permanent marker. The filter is removed and placed on the stub and the stub is placed in a vacuum evaporator for coating at a vacuum of 5.0 x 10⁻⁵ torr. The filter is then removed and placed in a clean polycarbonate storage box for transfer. When examining the sample, randomly located areas are selected for higher magnification scanning. Specific areas are scanned, morphology recorded and a spectrum is collected. Representative micrographs and spectra are stored digitally.

RESULTS AND DISCUSSION

Drug identification and authentication

As a prerequisite for any formulation preformulation determination like solubility and melting point were determined and the drug Canagliflozin was found to

be soluble in ethyl alcohol, methanol and practically insoluble in water and 0.1N HCl. Melting point was found to be 107.2°C which was further confirmed by DSC. FTIR spectral data showed a characteristic conjugated low C=O peak around 1500 cm⁻¹ and free, intermolecular alcohols (-OH) from 3358 cm⁻¹ 3738 cm⁻¹ and C=O stretch at 1743 cm⁻¹. All these findings were matched with literature values and found concurrent.

Drug polymer compatibility studies

The results obtained from preformulation compatibility studies (Figure 1 and 2) revealed that all the major peaks for drug and polymer were retained in FTIR spectrum and there were no significant changes in melting point of the drug (around 109°C) in physical mixture as seen in DSC thermographs. The results infer that the drug Canagliflozin is compatible with polymer.

Rheological properties

Flow property of liquid formulations is the major concern for pour ability and swallowing. Hence viscosities of the formulations were determined at 50, 75, 100 rpm

to get correspondent angular velocities and tabulated in Table 2. The viscosity of the formulations increased with an increase in sodium alginate concentration from 0.5 to 1.5% w/v. This phenomenon is as a consequence of increasing chain interaction with increase in polymer concentration. The calcium carbonate, which is the source of cation, when added decreased the viscosity of the formulation.

Buoyancy studies

The ideal gastroretentive system must have least floating lag time. The floating lag time of all the formulations were determined and values ranges from 157 s to 541 s. Formulation SF6 has the least floating lag time which consists of 1% sodium alginate and 1.5% calcium carbonate. The floating lag time of SF4 and SF5 were 286 s and 191 s respectively. Increase in floating lag time could be due to less concentration of calcium carbonate compared to SF6. This could be due to the volume of gas generated was high in the formulation SF6 and also the increase in concentration of cations in the formulation facilitates the fast gelation which is responsible for less floating lag time. Floating duration of all the formula-

Table 1: Formulation Table.

Sl. No.	Formulation Code	Polymer Concentration (%w/v)	Calcium Carbonate (%w/v)	Canagliflozin (%w/v)
1	SF1	1.5	0.5	1.0
2	SF2	1.5	1.0	1.0
3	SF3	1.5	1.5	1.0
4	SF4	1.0	0.5	1.0
5	SF5	1.0	1.0	1.0
6	SF6	1.0	1.5	1.0
7	SF7	0.5	0.5	1.0
8	SF8	0.5	1.0	1.0
9	SF9	0.5	1.5	1.0

Table 2: Evaluation parameters for formulations.

Sl. No.	Formulation	Viscosity at rpm in mPs			Drug Content (%)±SD (n=3)	FLT in Seconds±SD (n=3)
		50	75	100		
1	SF1	1009	991	754	100.47±0.15	270.00±8.69
2	SF2	1000	886	689	101.71±0.35	223.83±6.01
3	SF3	1102	1000	754	98.98±0.26	173.83±6.85
4	SF4	947	824	682	100.39±0.43	286.50±3.78
5	SF5	884	752	625	99.61±0.26	191.00±3.16
6	SF6	758	627	524	101.02±2.35	157.00±5.44
7	SF7	326	244	208	99.13±0.07	259.17±7.41
8	SF8	812	441	379	99.84±0.40	421.17±29.41
9	SF9	326	241	102	99.92±0.50	541.83±17.23

Table 3: In-vitro drug release data for GRDDS formulations.

SI No	Time in h	Cumulative in-vitro drug release for formulations (%) \pm SD (n=3)								
		SF1	SF2	SF3	SF4	SF5	SF6	SF7	SF8	SF9
1	0.25	17.82 \pm 0.016	18.52 \pm 0.004	16.32 \pm 0.016	17.98 \pm 0.004	20.13 \pm 0.005	7.51 \pm 0.005	29.35 \pm 0.013	15.24 \pm 0.007	16.52 \pm 0.007
2	0.5	18.94 \pm 0.011	20.34 \pm 0.007	19.45 \pm 0.011	18.98 \pm 0.002	26.35 \pm 0.026	11.23 \pm 0.008	35.15 \pm 0.009	23.14 \pm 0.003	22.13 \pm 0.018
3	0.75	20.64 \pm 0.040	25.22 \pm 0.011	21.68 \pm 0.003	25.42 \pm 0.008	32.41 \pm 0.006	13.74 \pm 0.064	41.71 \pm 0.005	31.05 \pm 0.004	41.20 \pm 0.021
4	1	22.01 \pm 0.016	28.97 \pm 0.009	23.84 \pm 0.005	26.12 \pm 0.007	33.26 \pm 0.017	16.58 \pm 0.013	43.33 \pm 0.008	46.02 \pm 0.023	43.45 \pm 0.009
5	2	33.57 \pm 0.012	31.25 \pm 0.008	33.25 \pm 0.015	37.42 \pm 0.010	45.27 \pm 0.007	34.27 \pm 0.010	62.17 \pm 0.024	71.96 \pm 0.015	58.25 \pm 0.004
6	3	35.24 \pm 0.007	33.28 \pm 0.004	39.08 \pm 0.012	47.52 \pm 0.005	52.13 \pm 0.015	38.67 \pm 0.020	68.28 \pm 0.023	78.25 \pm 0.016	67.24 \pm 0.027
7	4	39.24 \pm 0.003	37.24 \pm 0.015	40.68 \pm 0.008	51.26 \pm 0.006	56.87 \pm 0.006	40.21 \pm 0.019	82.84 \pm 0.025	93.01 \pm 0.005	73.27 \pm 0.015
8	5	44.52 \pm 0.009	46.78 \pm 0.011	44.27 \pm 0.006	54.25 \pm 0.012	59.23 \pm 0.005	43.28 \pm 0.006	95.24 \pm 0.027	96.08 \pm 0.007	78.05 \pm 0.017
9	6	49.42 \pm 0.017	49.21 \pm 0.009	48.03 \pm 0.003	56.28 \pm 0.013	60.21 \pm 0.038	47.21 \pm 0.003			80.21 \pm 0.016
10	7	53.24 \pm 0.194	51.56 \pm 0.018	48.75 \pm 0.009	57.21 \pm 0.026	63.24 \pm 0.009	56.53 \pm 0.002			84.25 \pm 0.010
11	8	61.48 \pm 0.011	54.21 \pm 0.008	50.23 \pm 0.021	63.08 \pm 0.024	68.23 \pm 0.016	60.28 \pm 0.024			92.08 \pm 0.011

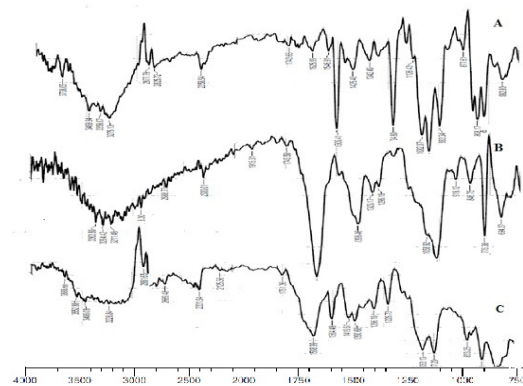


Figure 1: FTIR spectra for Canagliflozin Hemihydrate (A), Sodium Alginate (B) and Physical mixture (C).

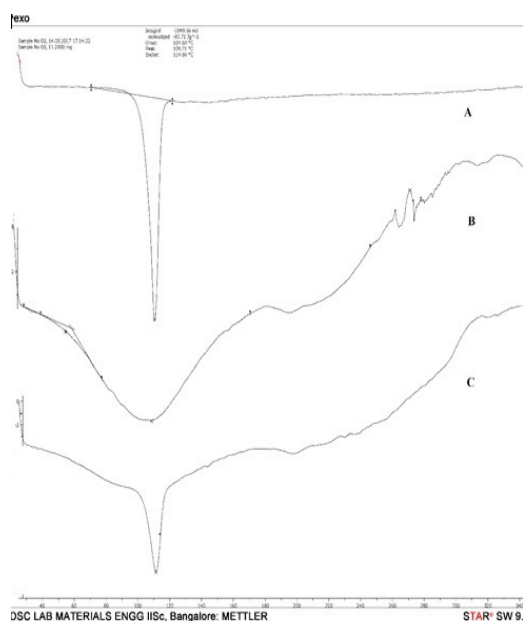


Figure 2: DSC thermogram for Canagliflozin Hemihydrate (A), Sodium Alginate (B) and Physical Mixture (C).

tions were more than 24 h in all the concentrations of sodium alginate and calcium carbonate.

In-vitro drug release study

All the formulations were subjected to *in-vitro* drug release and cumulative drug release versus time plot was depicted in Figure 3. The results of drug release suggested that at 15th m the drug release from formulations SF5 was 20.13% and that for SF4 and SF6 was 17.98 and 7.51% respectively. The reduction of drug release from SF6 could be due to quick conversion of liquid into gel. This observation were supported by floating lag time of 157 s for SF6 when compared with floating lag time of 191 s and 286 s for SF4 and SF5 respectively, where the amount of drug release from these two formulations comparatively higher than SF6 and floating lag time more than SF6. This observation also sug-

Table 4: Release Kinetics Data for Formulations.

Sl. No.	Formulation Code	Zero order R^2	First Order R^2	Higuchi R^2	Hixson R^2	Korsemeyer-Peppas	
						R^2	n -value
1	SF1	0.9102	0.9424	0.9958	0.9751	0.9874	0.4801
2	SF2	0.8134	0.8757	0.9740	0.8558	0.9758	0.3669
3	SF3	0.8001	0.9457	0.9046	0.9733	0.991	0.3490
4	SF4	0.8618	0.9198	0.9863	0.9021	0.9705	0.4876
5	SF5	0.8083	0.9578	0.9688	0.9371	0.9468	0.4166
6	SF6	0.8078	0.8735	0.9530	0.8532	0.948	0.4045
7	SF7	0.8385	0.9104	0.9829	0.8882	0.9776	0.4066
8	SF8	0.8454	0.8924	0.964	0.8772	0.9562	0.4200
9	SF9	0.7809	0.8709	0.9643	0.8426	0.9876	0.3350

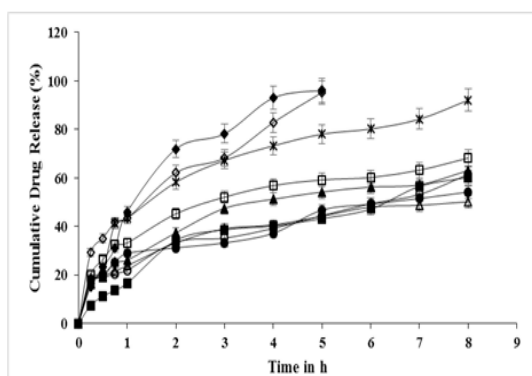


Figure 3: In-vitro drug release for formulations SF1[○], SF2[●], SF3[△], SF4[▲], SF5[□], SF6[■], SF7[◇], SF8[◆] and SF9^{*}.

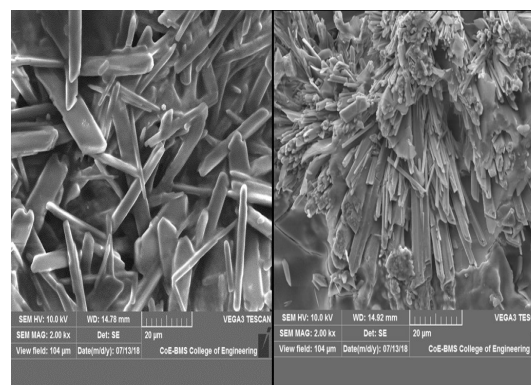


Figure 4: Scanning Electron Microscopic micrograms of SF5 formulation at 3 h and 5 h dissolution given in 2.00kx magnification.

gested that formation of gel was taken more time than SF6. The same trend continued for these formulations up to 8 h of drug release. Scanning electronic micrographs of gel formed at 3rd h and 5th h from SF5 formulation were studied and revealed that the formation of highly porous gel (Figure 4). This could be the reason for higher amount of drug release from SF5. The same pattern of drug release was found in SF1, SF2 and SF3. Among the formulations SF7, SF8 and SF9, in SF7 the amount of drug release at 15th m was 29.35 %, SF8 15.24 % and SF9 16.53 %. The increase in amount of drug release from SF7 suggested that the concentration of calcium carbonate was not enough to convert liquid form to gel and it forms a highly porous gel through which the drug was released. In the concentration of 1% and 1.5% of calcium carbonate stiff gel was formed there by it reduced the amount of drug release at 15th m. This observation was supported by floating lag times 259 s, 421 s and 547 s for SF7, SF8 and SF9 respectively. All these results of drug release and floating lag time of all the formulations suggested the influence of calcium carbonate on gelling properties of sodium alginate. These findings were correlated with the software assisted analysis of the responses (FLT, Viscosity, % *in-vitro* drug release at 1 h and 8 h) with respect to the

two different factors namely polymer concentration (A) and calcium carbonate concentration (B) at 3 different levels. (-1, 0, +1) (Table 5). Fit analysis, ANOVA and surface response 3D plots (Figure 5) for different responses were studied in detail. Fit analysis suggested linear model irrespective of response factors. Equations given as follows represent the effects of each factor on corresponding response, where A denotes sodium alginate concentration and B denotes calcium carbonate concentration.

$$\begin{aligned} \text{Floating Lag Time} &= +280.48 -92.42 A +9.50 B \\ \text{Viscosity} &= +667.33 +325.17 A -31.83 B \\ \text{Drug Release at 1 h} &= +30.50 -9.73 A -1.34 B \\ \text{Drug Release at 8 h} &= +70.04 -19.75 A -3.21 B \end{aligned}$$

For all the responses the model suggested is the linear model, ANOVA results were significant with *p*-values less than 0.05. The predicted and adjusted R^2 values are in reasonable agreement with each other. Adequate precision, i.e., signal to noise ratio is greater than 4 which is desirable. After setting the floating lag time, viscosity of the formulation to minimum possible values, release at 1

Table 5: Optimization using Design Expert Software.

Std	Run	Polymer	Gas generating agent	FLT in	Viscosity	%CDR	%CDR	%CDR	%CDR
		Concentration (%) (X1)	Concentration (%) (X2)	Seconds (Y1)	in mPs (Y2)	at 1 h (Y3)	at 3 h	at 5 h	at 8 h (Y4)
6	1	1.5	1	223.83	886	28.97	33.28	46.78	54.21
9	2	1	1	191.00	752	32.41	52.13	59.23	68.23
8	3	1	1.5	157.00	627	16.58	38.67	43.28	60.28
3	4	0.5	1.5	541.83	241	43.45	67.24	78.05	92.08
5	5	0.5	1	421.16	441	46.02	78.25	95.21	96.08
4	6	1.5	1.5	173.83	1000	22.01	35.24	44.52	61.48
2	7	1.5	0.5	270.00	991	23.84	39.08	44.27	50.23
1	8	0.5	0.5	259.16	244	43.33	68.28	95.24	95.24
7	9	1	0.5	286.50	824	26.12	47.52	54.25	63.08

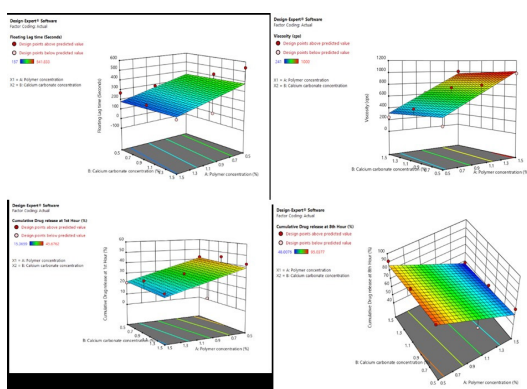


Figure 5: Three dimensional plots of formulation responses FLT, viscosity, % cumulative drug release at 1 h and 8 h for SF5.

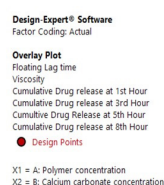


Figure 6: Graphical representation of optimization with flag stating optimized formulation parameters.

h to 30 % and release at 8 h to 90 % to attain the desired goals the search for the optimized formulation was carried out with the help of numerical optimization. The optimized formulation contained 0.868% sodium alginate and 0.957 % calcium carbonate to get floating lag time of 304.03 s, viscosity of 584.31 cps, drug release at 1 h 33.18% and 8 h 75.52 % respectively suggested by software. Considering the concentration of sodium alginate and calcium carbonate by the software was nearly matched with SF5 formulation. Graphical optimization chart with design space showing the optimized region with optimized formula is depicted in Figure 6.

Mechanism of drug release

Canagliflozin release from the formulations showed linearity towards Higuchi’s square root model, indicating the release mechanism to be diffusion based. R² values varied from 0.9046 for SF3 to 0.9958 for SF1. Results were tabulated in Table 4. The release exponents (n) of the formulations from Korsmeyer-Peppas model suggested that depending upon the formulation variables

the Canagliflozin followed Fickian mechanism (n=0.5 means Fickian diffusion, 0.5>n<1.0 non-Fickian diffusion and n=1.0 Case II diffusion). n-values varied from 0.3350 for SF9 to 0.4876 for SF4 (n<0.5). This could be attributed to the swelling behavior of the polymer in dissolution medium creating a barrier through which the drug diffuses as the time progresses.

CONCLUSION

In the present study by considering the effect of calcium cations on the gelling properties of various concentration of sodium alginate, it was concluded that the effective gelling was achieved in the formulation comprised of 1 % sodium alginate with 0.5 %, 1.0 % and 1.5 % of calcium carbonate. The software also suggested the optimized formulation must consists of 0.868 % of sodium alginate and 0.957 % of calcium carbonate and these compositions are much near to the composition of SF5 and thus SF5 was considered as optimized formulation.

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CONFLICT OF INTEREST

The author declare no conflict of interest.

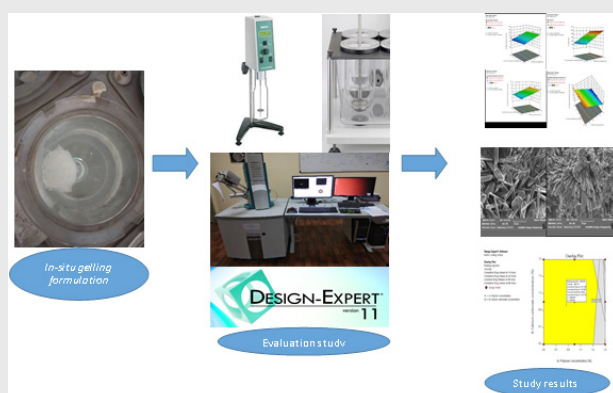
ABBREVIATIONS

GRT: Gastric Retention Time; **GET:** Gastric Emptying Time; **GRDDS:** Gastro Retentive Drug Delivery Systems; **FDSDS:** Floating Drug Delivery Systems; **HBS:** Hydrodynamically Balanced Systems; **GI:** Gastro Intestinal; **FTIR:** Fourier-Transform Infra-Red spectroscopy; **DSC:** Differential Scanning Calorimetry; **RPM:** Rotations Per Minute; **RP-HPLC:** Reverse Phase High Performance Liquid Chromatography; **CDR:** Cumulative Drug Release; **SEM:** Scanning Electronic Microscopy; **ANOVA:** Analysis of Variance.

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PICTORIAL ABSTRACT



SUMMARY

- In the current study an attempt was made to develop and optimize *in-situ* gelling liquid oral formulation of a novel antidiabetic drug; canagliflozin for the treatment of diabetes. The formulations were developed with different concentrations of sodium alginate and calcium carbonate. The developed formulations were evaluated for preformulation studies by FTIR and DSC, rheology, buoyancy and *in-vitro* drug release. A two-level three-response surface face centered central composite statistical experimental design for experimentation was carried out utilizing DESIGN EXPERT® version 11.0. Response surface graphs were used to show the factor interaction between the considered variables.
- By considering the effect of calcium cations on the gelling properties of various concentration of sodium alginate, it was concluded that the effective gelling was achieved in the formulation comprised of 1% sodium alginate with 0.5 %, 1.0 % and 1.5 % of calcium carbonate. The software also suggested nearly the composition of SF5 as optimized formulation.

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