

Neuroprotective Role of Periplocin Against Aluminium Chloride-stimulated Alzheimer's Disease in a Rat Model by Modulation of Oxidative Stress and Inflammation

Danni Li¹, Xiaoxiao Man¹, Xiaoxia Ma², Jinbo Sun^{1,*}

¹Department of Neurology, Central Hospital (Affiliated to Shandong First Medical University), Jinan, Shandong, CHINA.

²Digestive Endoscopy Center, Central Hospital (Affiliated to Shandong First Medical University), Jinan, Shandong, CHINA.

ABSTRACT

Background: Alzheimer's disease is a prevalent neurodegenerative condition marked by the assembly of β -amyloid deposits inside the brain parenchyma. Aluminium is a neurotoxin molecule that generates oxidative stress linked to various neurological disorders. **Objectives:** This study focused on the neuroprotective effects of periplocin, a natural compound, against aluminium chloride ($AlCl_3$)-stimulated diseased rat model. **Materials and Methods:** Four groups of twenty-four Sprague Dawley rats were established. For four weeks, the rats in the control group I was administered NaCl. $AlCl_3$ was provided to the second group of rats. An hour before $AlCl_3$ induction, rats in groups III and IV were given periplocin (25 and 50 mg/kg) orally. The rats were behaviourally examined before euthanization and analysis of the biochemical parameters was carried out after their sacrifice. **Results:** The learning ability and memory were impaired, lipid oxidation (MDA) was suppressed by antioxidant regulation (Reduced glutathione, Catalase, and Superoxide dismutase), and lactate dehydrogenase, $Na^+ K^+$ ATPase, acetylcholinesterase, and nitric oxide activity was inhibited upon periplocin administration. Additionally, the concentrations of inflammatory regulators such as $IL-1\beta$, $TNF-\alpha$, and $IL-6$ were suppressed. **Conclusion:** Thus, periplocin might be a crucial neuroprotector against the advancement of Alzheimer's disease.

Keywords: Periplocin, Alzheimer's disease, Aluminium chloride, Neuroinflammation, Acetylcholinesterase.

Correspondence

Dr. Jinbo Sun

Department of Neurology, Central Hospital (Affiliated to Shandong First Medical University), Jinan, Shandong, 250031, CHINA.
Email id: sunjb133610@outlook.com
ORCID ID 0000-0002-9029-2187

Received: 21-06-2022;

Revised: 24-09-2022;

Accepted: 11-10-2022.

INTRODUCTION

Alzheimer's Disease (AD) is a widespread neurological disorder that is marked by gradual cognitive deterioration, dementia, and memory loss that affects predominantly older individuals across the world.¹ Loss of neurons, amyloid plaques, and neurofibrillary tangles are all characteristics of AD.² Around 50 million people are reported to have been diagnosed with Alzheimer's disease worldwide, with the number anticipated to rise to 152 million by 2050. The disease is currently the 5th highest reason for mortality in adults of 65 years of age, and the 6th among people of all age categories.³

In the progression of AD, parts of the brain linked with memory loss and cognitive dysfunction, such as the cortical and hippocampal regions, are most impacted.⁴ Several behavioural facets of human existence including memory, critical thinking,

navigation, learning capacity and speed, apprehension, calculation, and judgment are considered to get affected by AD.⁵ Increases in glutamate concentration, neuroinflammation, and oxidative stress have all been connected with AD pathogenesis.⁶ Considering the several risk factors linked to this disease, it is difficult to understand its multifaceted pathophysiology and clinical aspects.⁷

The metal ion equilibrium in the brain is necessary for optimal cognitive functions, and its disruption has been recognized as one of the primary issues that contribute to neurodegenerative advancement.⁴ Among the environmental factors contributing to the risk of AD, exposure to metals such as aluminium is one of the widely known factors. By stimulating the immune system and releasing different inflammatory mediators, aluminium exposure causes immuno-excitotoxicity, which leads to neurotoxicity.⁸

Al has been proven to penetrate the blood-brain barrier (BBB) via a particular transferrin receptor and cause severe loss of memory by disrupting a variety of typical neuronal functioning. Aluminium can serve as a cross-linker for amyloid protein, causing it to oligomerize and cause neurotoxicity.⁹



DOI: 10.5530/001954642194

Copyright Information

© 2023 Author(s). Distributed under Creative Commons CC-BY 4.0

Publishing partner : EManuscript Tech [www.emanuscript.in]

Current treatment for Alzheimer's disease only gives relief from the symptoms with significant adverse effects that can worsen to stroke as well as even death.¹⁰ As a result, new medications for treating or managing AD are urgently required. The focus of worldwide therapeutic research is now focusing on herbal medicine. Herbal plants have long been employed as traditional medicine since they are inexpensive and have few adverse effects. Numerous bioactive chemicals isolated from plants have been shown to minimize neuronal injuries in recent years, and they have become an important treatment alternative for related brain diseases.^{4,11}

Cortex periplocae is herbal medicine employed traditionally over the years that possesses immune system enrichment, anticancer, antiradiation, and anti-inflammatory properties.¹² Periplocin is a cardiac glycoside identified and isolated from the plant that is utilized to treat rheumatoid arthritis and bone and tendon strengthening.¹³ It also has a powerful inhibitory impact on the propagation of various cancer cell lines like colon, lung, and gastric cancer.¹⁴ Even though the effectiveness of periplocin as an anti-inflammatory drug in brain barrier tissues is yet to be determined, cardiac glycosides often possess a wide array of chemical variety as well as absorption, distribution, metabolism, elimination, and toxicity characteristics.¹⁵

With this background, the primary purpose of the current study is to assess the neuroprotective potential of periplocin against AlCl₃-stimulated oxidative damage, cognitive decline, transmembrane activity, and neuroinflammation in Sprague Dawley (SD) rats.

MATERIALS AND METHODS

Reagents and chemicals

For this study, periplocin from an analytical standard was bought. AlCl₃ was prepared in 0.9 percent saline, BCA protein assay kit, antioxidant ELISA kit Griess reagent, RNase, and Trizol for RNA isolation, Lactate dehydrogenase (LDH) detection kit, dinucleotide triphosphates, Taq polymerase, and reverse transcriptase enzyme were all purchased from Sigma-Aldrich. The other chemicals employed in the study were of analytical quality.

Animal model

Sprague Dawley rats of age 7–9 weeks and weight 170–190 g, were caged and provided tap water and pellets in a contained environment. Before the experiment, the animals were familiarized with the experimental environment for 7 days in the animal housing. The procedures were carried out to ensure that the experimental rats suffered as little as possible, and ethical approval for the same was obtained by the Institutional Animal Ethics Committee (IAEC).

Experimental design

The experimental animals were categorized into treatment groups before starting the experiment. As a result, the rats were separated into 4 groups, each with six rats. The rat's body weight was taken into account while administering the dose (B.W.). The animals in group I were given 0.1 percent NaCl for the duration of the trial (4 weeks) and were considered the control group. The rats in group II were administered AlCl₃ (100 mg/kg b.wt) dissolved in water. Periplocin (25 mg/kg and 50 mg/kg b.wt) was given orally to rats in groups III-IV, one hour before AlCl₃ stimulation. The animal groupings for various treatments are as follows: Group I – Control (0.1% NaCl) for 4 weeks; Group II – 500 ul of AlCl₃ (100 mg/kg b.wt) for 4 weeks; Group III – Periplocin (25 mg/kg b.wt) treatment 1h before AlCl₃ (100 mg/kg b.wt) exposure for 4 weeks; Group IV – Periplocin (50 mg/kg b.wt) treatment 1h before AlCl₃ (100 mg/kg b.wt) exposure for 4 weeks.

Food ingestion and weight fluctuations in the rodents were monitored daily. Before being sacrificed, all of the rats were put through behavioural testing. Pentobarbital sodium (40 mg/kg) was utilized to anesthetize the rats, and brain and blood samples were rapidly collected for biochemical investigation.

Estimation of behavioural indices

To study the memory and learning potential of rodent models, behavioural investigations employing the maze test were conducted. To perform the tasks in the rats, the RAM and MWM tests were employed.

Radial arm maze (RAM) test

As per the protocol of Foyet *et al.*¹⁶ the RAM test was carried out with minor adjustments. Experimental rats were fed a rigorous diet to promote appetite during the study. In a fixed layout in the room, many visual extra-maze signals were supplied at a distance of 10–30 cm to the maze. At the termination of each arm, around feeding well (0.5 cm depth with a 1 cm diameter) was placed. The animals were stationed in the stage centre with feed pellets on the first day. The rats were then permitted to run to the baited arms for eating for the following five days. On the eighth day, the rats were subjected to the same training approach and were given reference and working memory exercises. The measurements to be recorded were the count of reference memory errors (RMEs) and the count of working memory errors (WMEs).

Morris water maze (MWM) test

The MWM assessment was carried out by following the protocol of Ahmed and Gilani with minimal modifications.¹⁷ For the investigation, a spherical stage with a puddle of water was established at the base. Before the treatment, the animals were permitted to swim from different places to the platform's end, and the duration it took them to get out of the water (Escape Latency-

ELT) was recorded. The rats' ELT to turn up at the platform after treatment was noted and plotted against the training day number.

Tissue homogenate

The brain sample was divided into 2 equal halves. One-half of the sample was preserved in saline (10%) for biochemical investigation, while the other portion was preserved in formaldehyde (4%) for histological investigation. The tissues were subjected to homogenization in PBS solution and centrifuged for 20 min at maximum speed (4°C) for biochemical investigation. The protein concentrations in the resultant supernatant were estimated with the BCA protein assay kit.

Estimation of oxidative-antioxidative indicators

Colorimetric and antioxidant ELISA test kits were utilized to analyse the homogenate oxidative stress marker, i.e., Malondialdehyde along with the antioxidant markers, i.e., glutathione, catalase, and superoxide dismutase (SOD), respectively. The complete protocols were carried out as per the manufacturer's guidelines.

Malondialdehyde (MDA)

The protocol was provided by Jain *et al.*¹⁸ was utilized to estimate the tissue MDA concentration. PBS, BHT, and 30% TC were combined with the homogenate solution. The reaction mix was subjected to centrifugation (20 min, 3000 Xg) after a 1hr incubation at 37°C. After that, EDTA and 1% TBA were added to the supernatant. For at least 20 min, this mix was left for incubation at 80°C in a water bath. MDA was measured at 532 nm and represented as nmol/g of wet tissue.

Superoxide dismutase (SOD)

The SOD enzyme was calculated using the approach of Marklund and Marklund.¹⁹ The concentration of SOD was primarily assessed by its capacity to block superoxide-related decrease. The quantity of enzyme necessary to combat pyrogallol oxidation by 50% was considered to be one unit of SOD. SOD activity was measured in units/mg of protein.

Catalase (CAT)

The activity of CAT was estimated using Aebi's technique.²⁰ The quantity of enzyme required to breakdown 1mol of hydrogen peroxide (H₂O₂) per minute was defined as one CAT unit. The CAT enzyme activity was measured in units per milligram of protein.

Reduced glutathione (GSH)

The GSH level was determined using the Aykac *et al.* technique.²¹ The reaction solution contained a combination of homogenate supernatant (2 ml), Millipore water (8 ml), and 1M NaCl (100

ml, pH 4.7). The activity was measured in nmol/mg protein and the absorbance was determined at 400 nm.

Estimation of lactate dehydrogenase (LDH)

An LDH toxicity detection kit was used to measure periplocin's tissue damage activity. In 0.1 M of K₃PO₄ buffer, the supernatant of homogenate was combined with Triton X-100 (6%, 40 µl) and 4.6 mM pyruvic acid (100 µl, pH 7.5). Using an ELISA microplate reader, the absorbance was determined at 340 nm. With the equation given below, the LDH release was calculated as: LDH leakage (%) = [A] sample / [A] control x 100; [A] Sample: Sample absorbance value. [A] Control: Control absorbance value.

Estimation of nitric oxide (NO) in AlCl₃ triggered rats

The concentrations of NO were measured using the Griess reagent, as described by Tracey *et al.*²² After dilution with PBS, the tissue homogenates were plated on a 96-well plate and left for incubation at 25°C for 15 min. A spectrophotometer was employed to measure the absorbance value against a blank at 540 nm.

Measurement of acetylcholinesterase (AChE) enzyme activity

Ellman's approach²³ was adopted to determine the AChE activity. Tris-HCl (50 mM, 170 ml), supernatant (25 and 50 mg/kg, 250 ml), AChE (10 ml, 6.67 U ml⁻¹), and DTNB (5,5'-dithio-bis [2-nitrobenzoic acid]) (20 ml, 10 mM) in buffer were combined. After that, the reaction solution was incubated for 10 min at 37°C. The absorbance was determined at 412 nm after the addition of acetylthiocholine iodide (200 mM, 10 µl). The percentage of enzyme inhibition was estimated as follows: Inhibition percentage = 100 - Change of Absorbance of test/Change of Absorbance of blank × 100

Estimation of the transmembrane protein activity

The activity of the Na⁺/K⁺ ATPase was evaluated by following the protocol of Li *et al.*²⁴ The homogenate supernatant was left for incubation at 37°C in a mix of 0.2 ml each of buffer, distilled water, calcium chloride, and ATP. The mixture was then incubated for another 10 min at 37°C with 20% TCA (2 ml). Upon addition of ammonium molybdate (1 ml) and 4-aminonaphthol sulphonic acid (0.8 ml), a blue tint appeared. The resultant mixture's absorbance was analyzed at 650 nm.

Estimation of inflammatory cytokines in AlCl₃ triggered rats

ELISA was utilized to study the levels of inflammation-associated cytokines, IL-1β, IL-6, and TNF-α. We performed ELISA tests on the diluted brain tissue extract to assess the concentrations of IL-1β (ab100768), IL-6 (ab100772), and TNF-α (ab100785) using abcam ELISA kits (abcam, USA) according to the manufacturer's instructions. The detection limits of each were 68.59 pg/ml -

50000 pg/ml for IL-1 β , 40.96 pg/ml - 10000 pg/ml for IL-6, and 82.3 pg/ml - 20000 pg/ml for TNF- α .

Statistical analysis

The results were calculated with SPSS software (version 17.0) and displayed as the mean \pm SD of triplicate trials. ANOVA was used to compare the experimental groups, followed by the Student-Newman-Keuls multiple comparison test. Statistical significance was assessed at $p < 0.05$.

RESULTS

Impact of Periplocin treatment on behavioural parameters in AlCl₃ provoked animals

On the behavioral parameters of rats stimulated with AlCl₃ and treated with periplocin, the mean time taken to complete the maze was measured and compared between groups (Figure 1). The impact of Periplocin treatment on RMEs, WMEs, and TL time in the control and treated groups are depicted in Figure 2. In

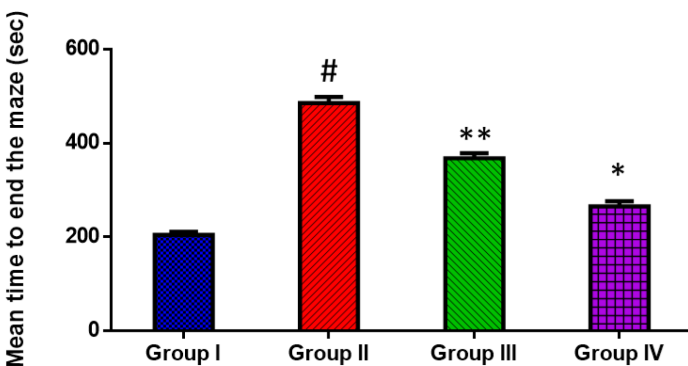


Figure 1: The mean total time to complete the maze in the periplocin treatment in AlCl₃-stimulated rats. Periplocin was administered (25 and 50 mg/kg) to the rats in order to measure behavioural parameters and determine total time spent in the maze. The data are represented as Mean \pm SD (* $p < 0.05$; ** $p < 0.005$, # $p < 0.001$).

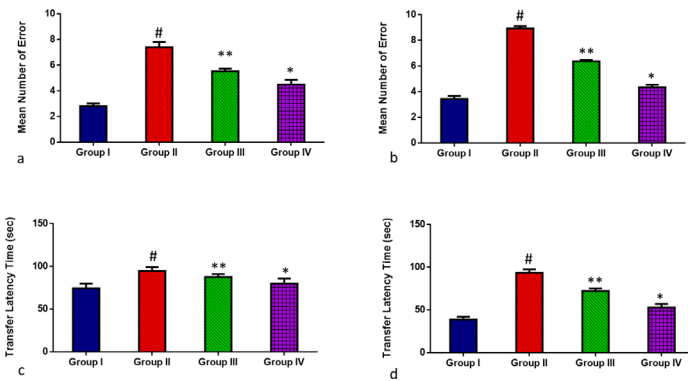


Figure 2: The impact of periplocin treatment on the behavioral parameters in AlCl₃-stimulated rats. The change in reference memory errors (a), working memory errors (b), initial transfer latency time (c), and Retention transfer latencies time (d) by periplocin administration (25 and 50 mg/kg) are given. The data are represented as Mean \pm SD (* $p < 0.05$; ** $p < 0.005$, # $p < 0.001$).

comparison with the group I animals, the RMEs, WMEs, and TL values of AlCl₃ induced rats were considerably higher to perform the tasks. When compared to AlCl₃-triggered rats, periplocin treatments at 25 mg/kg and 50 mg/kg resulted in a substantial decrease in all variables.

Impact of Periplocin administration on oxidative stress levels in AlCl₃ provoked animals

The impact of Periplocin treatment on oxidative stress levels was estimated by determining the MDA and antioxidant enzyme activities in brain samples, such as CAT, GSH, and SOD in the control and treated samples (Figure 3). The MDA levels of AlCl₃ administered to animals were substantially greater than in control rats, whereas Periplocin (25 and 50 mg/kg) administration significantly reduced the MDA level (Figure 3a). In contrast, the CAT, GSH, and SOD levels in the AlCl₃ administrated rats were considerably lesser than in the control samples (Figure 3b-3d). The findings may propose periplocin's remarkable antioxidant alleviation impact on AlCl₃-stimulated oxidative stress in rodents.

Impact of Periplocin administration on neuronal damage in AlCl₃ provoked animals

Figures 4a and 4b demonstrate the effect of Periplocin administration on neuronal damage as measured by LDH and NO activities in the tissue samples of the control and treated groups. When comparing the AlCl₃ induced group with the control rats, the data revealed that LDH and NO activity were considerably higher in the AlCl₃ induced animals. In comparison with the AlCl₃ induced animals, rodents treated with Periplocin at both concentrations demonstrated a reversed attenuated impact of LDH and NO activity with a notable difference.

Impact of Periplocin administration on cholinergic functioning in AlCl₃ provoked animals

The activity of Na⁺ K⁺ ATPase and AchE in the tissue samples of control and treated groups is shown in Figure 5 to evaluate the

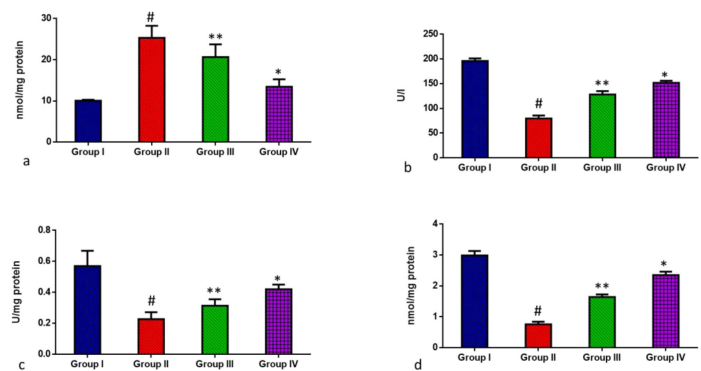


Figure 3: The oxidative stress level of periplocin in AlCl₃-stimulated rats. The change in markers Malondialdehyde (a), Superoxide dismutase (b), Catalase (c), and Glutathione (d) by periplocin administration (25 and 50 mg/kg) are shown. The data are represented as (* $p < 0.05$; ** $p < 0.005$, # $p < 0.001$).

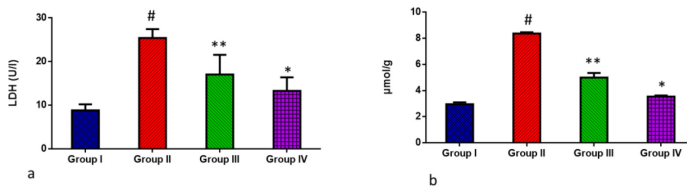


Figure 4: The neuronal damage effects of periplocin in $AlCl_3$ -stimulated rats. The activities of lactate dehydrogenase (a) and Nitrite Oxide (b) by periplocin administration (25 and 50 mg/kg) are depicted. The data are represented as Mean \pm SD (* $p < 0.05$; ** $p < 0.005$, # $p < 0.001$)

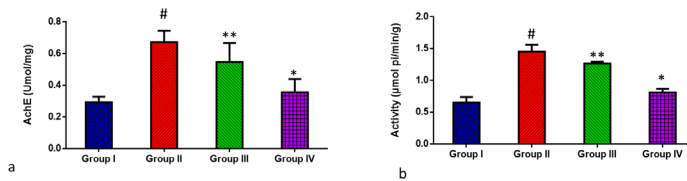


Figure 5: The impact of periplocin administration on cholinergic functioning in $AlCl_3$ provoked animals. The activities of acetylcholinesterase (a) and $Na^+ K^+$ ATPase (b) by periplocin administration (25 and 50 mg/kg) are illustrated. The data are represented as Mean \pm SD (* $p < 0.05$; ** $p < 0.005$, # $p < 0.001$).

influence of Periplocin administration on cholinergic function. The levels of $Na^+ K^+$ ATPase as well as AchE were increased in tandem in animals from $AlCl_3$ provoked and control group, as shown in Figure 5a and 5b, with a notable difference between both the groups. Following that, the AchE and $Na^+ K^+$ ATPase levels in rats were considerably decreased by Periplocin treatment (25 and 50 mg/kg). This is due to the increase in cholinergic functioning by periplocin administration, which is crucial for memory, learning, and motor function in the brain.

Impact of Periplocin administration on inflammatory cytokines expression in $AlCl_3$ provoked animals

The influence of Periplocin administration on the expression of inflammation-associated signalling pathways in control and treated animals is shown in Figure 6. In comparison with the control group, the levels of pro-inflammatory molecules such as TNF- α , IL-6, and IL-1 β exhibited a tremendous increase in the $AlCl_3$ provoked rats. In animals from Groups III and IV, however, Periplocin administration (25 and 50 mg/kg) dramatically reversed the $AlCl_3$ impact.

DISCUSSION

The major clinical sign of Alzheimer's disease has been identified as cognitive deficits along with serious health issues caused by Al exposure. Al-stimulated neurochemical and biochemical aberrations have been linked to cognitive deficits, and comparable abnormalities have been observed in Alzheimer's disease.²⁵ Many age-related problems and neurodegenerative diseases are driven by enhanced oxidative stress, therefore the search for natural medicinal agents that boost cognitive function and enable neuroprotective effects via antioxidant action is of

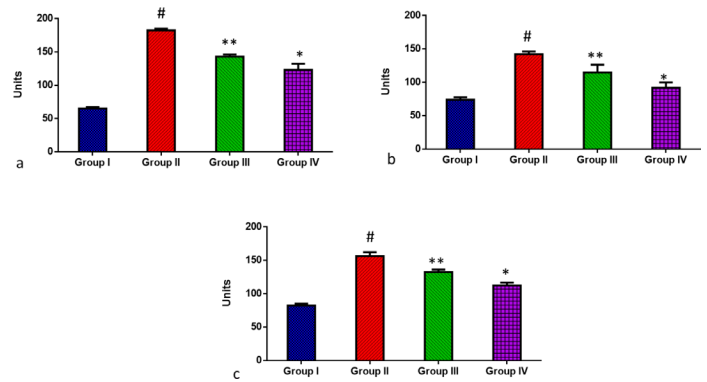


Figure 6: The impact of periplocin administration on inflammation-related mRNA gene expression in $AlCl_3$ provoked animals such as TNF- α (a), IL-6 (b) and IL-1 β (c). The gene estimation by periplocin administration (25 and 50 mg/kg) is shown. The data are represented as Mean \pm SD (* $p < 0.05$; ** $p < 0.005$, # $p < 0.001$).

significant interest.^{26,27} In the current investigation, periplocin was administered to $AlCl_3$ -induced rats to determine whether it might alleviate behavioural, biochemical, and neurochemical problems, as well as to evaluate its potential as a neuroprotective drug against AD.

An association between aluminium-induced neurotoxicity and AD pathogenesis has been proposed in prior studies.²⁸ Long-term exposure to aluminium has been shown to produce disorientation, memory problems, and eventually dementia due to the brain's poor clearance of aluminium.^{29,30} Increased AchE activity, oxidative stress, tau protein a reduction in BDNF, and aggregation of A β plaques are among the additional variables that mediate aluminium contribution in AD pathogenesis.³¹ This encouraged us to explore the effect of periplocin in an $AlCl_3$ -stimulated rat model. Aluminium induction hampered the animal's capacity to learn as well as remember the platform's position in both maze examinations, most likely owing to a hippocampal deficiency. Interestingly, periplocin administration inverted this impact and increased the rats' learning capabilities and spatial memory. Black pepper has been demonstrated to improve memory in the $AlCl_3$ stimulated neurotoxicity animal model in previous investigations.³²

The treatment of $AlCl_3$ resulted in a considerable increase in the duration necessary to complete the activity in the radial arm maze, indicating a reduction in working and reference memory.⁴ In rats, increased Al intake causes A β build-up in the cortical and hippocampal brain areas, resulting in memory and learning impairments.³³ The Morris maze was employed to assess spatial memory, and animals administered with $AlCl_3$ alone had a lower likelihood to recall the position of the hidden platform, even after being instructed over several weeks. The memory deficit caused by $AlCl_3$ was reversed when periplocin was administered, demonstrating an elevated memorizing capacity of periplocin.

An aged brain is susceptible to oxidative stress because of its high propensity for oxygen uptake, elevated iron content, polyunsaturated fatty acid concentration in the membranes, low glutathione levels, and limited antioxidant defence capabilities.³¹ Also, the brain is considered to be highly vulnerable to the negative impact of aluminium, and it is particularly susceptible to oxidative stress caused by excessive concentrations of free radicals along with inadequate levels of antioxidants.³⁴ As a result, the oxidant-antioxidant imbalance has been acknowledged as a crucial component in the progress of AD. Prolonged treatment of AlCl_3 led to elevated oxidative stress, as evidenced by a significant rise in lipid peroxidation as well as nitrite concentrations, along with a reduction in superoxide dismutase, reduced glutathione, and catalase levels in our study.

Aluminium-stimulated reduction in both axonal mitochondrial turnover and synaptic vesicles trigger oxidative metabolite release in neurons, which might explain the apparent increase in MDA levels and reduction in GSH concentration caused by AlCl_3 administration.³⁵ MDA and NO levels in living organisms are key markers of oxidative stress. MDA is formed when ROS induces peroxidation of membrane lipids, resulting in membrane damage and degradation. NO is a neurotransmitter in the central nervous system, yet it is neurotoxic when it is produced in excess.^{26,36} By clearing up the augmented ROS in the tissue samples, the combination therapy of AlCl_3 along with periplocin revived the oxidative stress indicators. Furthermore, elevated lipid peroxidation as a result of oxidative stress might explain the reduction in brain weight.³⁷

The pathophysiology of Alzheimer's disease is strongly linked to an uncontrolled neuroinflammatory process, like the overexpression of inflammatory mediators like $\text{IL-1}\beta$, which in turn suppresses the formation of the brain-derived neurotrophic factors responsible for the conduct of physiological functions in the CNS.^{38,39} Furthermore, patients with high levels of AlCl_3 demonstrated cognitive deterioration, which has been linked to impaired pro-inflammatory regulators including $\text{TNF-}\alpha$.³¹ Cardiac glycosides like periplocin may be beneficial in addressing inflammation at the brain's periphery, opening up new avenues for medication development for neuroinflammatory degeneration.²⁸ In the present investigation, the overproduction of $\text{TNF-}\alpha$, $\text{IL-1}\beta$, and IL-6 in the tissue homogenate of aluminium-intoxicated rodents treated with periplocin was significantly suppressed, which indicates its potent anti-inflammatory effect.

AchE is a broadly distributed particularly potent enzyme in the brain that has several roles in cholinergic and neuromuscular synapses and is linked with building and preserving learning memory in the brain.⁴⁰ Owing to the action of transmembrane protein $\text{Na}^+ \text{K}^+ \text{ATPase}$, AchE is responsible for maintaining the cholinergic membrane stability. Memory loss in AD is connected to the malfunction of cholinergic transmission.⁴¹ The treatment of AlCl_3 to rats led to a considerable elevation in AchE and

transmembrane protein activity in our investigation. Periplocin administration in the AlCl_3 -induced animals demonstrated a neuroprotective effect via inhibiting the AchE and transmembrane protein activity. Similarly, the suppressive action of a natural compound, fenugreek along with its active component on AchE has been discovered.⁴² The amount of acetylcholine is elevated when the AchE potential is reduced, which ultimately has positive implications on cognitive performance.²

CONCLUSION

Prolonged exposure to aluminium chloride may promote lipid peroxidation due to ROS production, resulting in increased ACHE activity and lower acetylcholine levels, impairing memory as well as cognition. In the present investigation, Periplocin was found to attenuate AlCl_3 -induced learning deficits, tissue damage, and cholinergic dysfunction via suppression of reactive free radicals and inflammatory mediators, owing to periplocin's powerful antioxidant and anti-inflammatory potential. These findings propose that Periplocin may be a possible therapeutic option for neurodegenerative diseases. However, before clinical trials, further research is required to confirm periplocin's neuroprotective activity in various Alzheimer's disease models.

ACKNOWLEDGEMENT

The authors would like to express their gratitude to Central Hospital (Affiliated to Shandong First Medical University), for providing the financial support to conduct this research.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ABBREVIATIONS

IAEC: Institutional Animal Ethics Committee; **RAM:** Radial arm maze; **MWM:** Morris water maze; **MDA:** Malondialdehyde; **SOD:** Superoxide dismutase; **GSH:** Reduced glutathione.

SUMMARY

- Periplocin is a cardiac glycoside identified and isolated from the *Cortex periplocae*.
- It exhibits Neuroprotective against aluminium chloride (AlCl_3)-stimulated diseased rat model.
- On treating the aluminium chloride (AlCl_3)-stimulated Alzheimer's disease with periplocin, the behavioral study and MDA level were significantly suppressed.
- Periplocin treatment suppressed lactate dehydrogenase, $\text{Na}^+ \text{K}^+ \text{ATPase}$, acetylcholinesterase, and nitric oxide activity as well as antioxidant regulation such as Reduced glutathione, Catalase, and Superoxide dismutase.

- The suppression of inflammatory regulators like IL-1 β , TNF- α , and IL-6 were observed on treatment with periplocin.

Author Contribution

The authors contributed equally.

Ethics Approval

All work has been done under the guidelines of the Institutional Ethics Committee.

Consent to Participate

All authors have their consent to participate.

Consent for Publication

All authors have their consent to publish their work.

REFERENCES

- Hajipour S, Sarkaki A, Farbood Y, Eidi A, Mortazavi P, Valizadeh Z. Effect of gallic acid on dementia type of Alzheimer disease in rats: Electrophysiological and histological studies. *Basic Clin. Neurosci J*. 2022 Jun;7(2):97-106. doi: 10.15412/J.BCN.03070203.
- Yang X, Du W, Zhang Y, Wang H, He M. Neuroprotective effects of higenamine against the Alzheimer's disease via amelioration of cognitive impairment, A β Burden, Apoptosis and Regulation of Akt/GSK3 β Signaling Pathway. *Dose-Response*. 2020;18(4):155932582097220:1559325820972205. doi: 10.1177/1559325820972205, PMID 33354171.
- Shunan D, Yu M, Guan H, Zhou Y. Neuroprotective effect of betalain against A β 1-42-induced Alzheimer's disease in Sprague Dawley Rats via putative modulation of oxidative stress and nuclear factor kappa B (NF- κ B) signaling pathway. *Biomed Pharmacother*. 2021;137:11369. doi: 10.1016/j.biopha.2021.111369, PMID 33582452.
- Justin Thenmozhi A, William Raja TR, Manivasagam T, Janakiraman U, Essa MM. Hesperidin ameliorates cognitive dysfunction, oxidative stress and apoptosis against aluminum chloride induced rat model of Alzheimer's disease. *Nutr Neurosci*. 2017;20(6):360-8. doi: 10.1080/1028415X.2016.1144846, PMID 26878879.
- Sanabria-Castro A, Alvarado-Echeverria I, Monge-Bonilla C. Molecular pathogenesis of Alzheimer's disease: an update. *Ann Neurosci*. 2017;24(1):46-54. doi: 10.1159/000464422, PMID 28588356.
- Wang R, Reddy PH. Role of glutamate and NMDA receptors in Alzheimer's disease. *J Alzheimers Dis*. 2017;57(4):1041-8. doi: 10.3233/JAD-160763, PMID 27662322.
- Moussa-Pacha NM, Abdin SM, Omar HA, Alniss H, Al-Tel TH. BACE1 inhibitors: current status and future directions in treating Alzheimer's disease. *Med Res Rev*. 2020;40(1):339-84. doi: 10.1002/med.21622, PMID 31347728.
- Chavali VD, Agarwal M, Vyas VK, Saxena B. Neuroprotective effects of ethyl pyruvate against aluminum chloride-induced Alzheimer's disease in rats via inhibiting toll-like receptor 4. *J Mol Neurosci*. 2020;70(6):836-50. doi: 10.1007/s12031-020-01489-9, PMID 32030557.
- Zaky A, Bassiouny A, Farghaly M, El-Sabaa BM. A combination of resveratrol and curcumin is effective against aluminum chloride-induced neuroinflammation in rats. *J Alzheimers Dis*. 2017;60(s1):S221-35. doi: 10.3233/JAD-161115, PMID 28222524.
- Liu KY, Stringer AE, Reeves SJ, Howard RJ. The neurochemistry of agitation in Alzheimer's disease: a systematic review. *Ageing Res Rev*. 2018;43:99-107. doi: 10.1016/j.arr.2018.03.003, PMID 29524596.
- Mohd Sairazi NS, Sirajudeen KNS. Natural products and their bioactive compounds: neuroprotective potentials against neurodegenerative diseases. *Evid Based Complement Alternat Med*. 2022 Jun;2020:1-30. doi: 10.1155/2020/6565396.
- Xie G, Sun L, Li Y, Chen B, Wang C. Periplocin inhibits the growth of pancreatic cancer by inducing apoptosis via AMPK-mTOR signaling. *Cancer Med*. 2021;10(1):325-36. doi: 10.1002/cam4.3611, PMID 33231372.
- Lu ZJ, Zhou Y, Song Q, Qin Z, Zhang H, Zhou YJ, et al. Periplocin inhibits growth of lung cancer *in vitro* and *in vivo* by blocking AKT/ERK signaling pathways. *Cell Physiol Biochem*. 2010;26(4-5):609-18. doi: 10.1159/000322328, PMID 21063098.
- Li L, Zhao LM, Dai SL, Cui WX, Lv HL, Chen L, et al. Periplocin extracted from cortex *Periplocace* induced apoptosis of gastric cancer cells via the ERK1/2-EGFR pathway. *Cell Physiol Biochem*. 2016;38(5):1939-51. doi: 10.1159/000445555, PMID 27160973.
- Botelho AFM, Piezezan F, Soto-Blanco B, Melo MM. A review of cardiac glycosides: structure, toxicokinetics, clinical signs, diagnosis and antineoplastic potential. *Toxicol*. 2019;158:63-8. doi: 10.1016/j.toxicol.2018.11.429, PMID 30529380.
- Foyet HS, Asongalem AE, Oben EK, Cioanca O, Hancianu M, Hritcu L. Effects of the methanolic extract of *Vitellaria paradoxa* Stem Bark against scopolamine-induced cognitive dysfunction and oxidative stress in the rat hippocampus. *Cell Mol Neurobiol*. 2016;36(7):1139-49. doi: 10.1007/s10571-015-0310-7, PMID 26620052.
- Ahmed T, Gilani AH. Inhibitory effect of curcuminoids on acetylcholinesterase activity and attenuation of scopolamine-induced amnesia may explain medicinal use of turmeric in Alzheimer's disease. *Pharmacol Biochem Behav*. 2009;91(4):554-9. doi: 10.1016/j.pbb.2008.09.010, PMID 18930076.
- Jain SK, McVie R, Duett J, Herbst JJ. Erythrocyte membrane lipid peroxidation and glycosylated hemoglobin in diabetes. *Diabetes*. 1989;38(12):1539-43. doi: 10.2337/diab.38.12.1539, PMID 2583378.
- Marklund S, Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur J Biochem*. 1974;47(3):469-74. doi: 10.1111/j.1432-1033.1974.tb03714.x, PMID 4215654.
- Aebi U, Fowler WE, Rew P, Sun TT. The fibrillar substructure of keratin filaments unraveled. *J Cell Biol*. 1983;97(4):1131-43. doi: 10.1083/jcb.97.4.1131, PMID 6194161.
- Aykaç G, Uysal M, Yalçın AS, Koçak-Toker N, Sivas A, Oz H. The effect of chronic ethanol ingestion on hepatic lipid peroxide, glutathione, glutathione peroxidase and glutathione transferase in rats. *Toxicology*. 1985;36(1):71-6. doi: 10.1016/0300-483x(85)90008-3, PMID 4040665.
- Tracey WR, Tse J, Carter G. Lipopolysaccharide-induced changes in plasma nitrite and nitrate concentrations in rats and mice: pharmacological evaluation of nitric oxide synthase inhibitors. *J Pharmacol Exp Ther*. 1995;272(3):1011-5. PMID 7534350.
- ELLMAN GL, COURTNEY KD, ANDRES V, FEATHER-STONE RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol*. 2022 Jun;7(2):88-95. doi: 10.1016/0006-2952(61)90145-9.
- Li Z, Li S, Hu L, Li F, Cheung AC, Shao W, et al. Mechanisms underlying action of xinmailong injection, a traditional Chinese medicine, in cardiac function improvement. *Afr J Tradit Complement Altern Med*. 2017;14(2):241-52. doi: 10.21010/ajtcam.v14i2.26, PMID 28573241.
- Cao Z, Wang F, Xiu C, Zhang J, Li Y. *Hypericum perforatum* extract attenuates behavioral, biochemical, and neurochemical abnormalities in aluminum chloride-induced Alzheimer's disease rats. *Biomed Pharmacother*. 2017;91:931-7. doi: 10.1016/j.biopha.2017.05.022, PMID 28514831.
- Abowlwafa HR, El-kott AF, Abd-Ella EM, Yousef HN. The possible neuroprotective effect of silymarin against aluminum chloride-prompted alzheimer's-like disease in rats. *Brain Sci*. 2020;10(9):628. doi: 10.3390/brainsci10090628, PMID 32932753.
- Mohamed NE-S, Abd El-Moneim AE. *Ginkgo biloba* extract alleviates oxidative stress and some neurotransmitters changes induced by aluminum chloride in rats. *Nutrition*. 2017;35:93-9. doi: 10.1016/j.nut.2016.10.012, PMID 28241996.
- Jansson D, Dieriks VB, Rustenhoven J, Smyth LCD, Scotter E, Aalderink M, et al. Cardiac glycosides target barrier inflammation of the vasculature, meninges and choroid plexus. *Commun Biol*. 2021;4(1):260. doi: 10.1038/s42003-021-01787-x, PMID 33637884.
- Yaseen AA, Al-Okbi SY, Hussein AMS, Mohamed DA, Mohammad AA, Fouda KA, et al. Potential protection from Alzheimer's disease by wheat germ and rice bran nanoform in rat model. *J App Pharm Sci*. 2022 Jun;9:67-76. doi: 10.7324/JAPS.2019.5108.
- Wenting L, Ping L, Haitao J, Meng Q, Xiaofei R. Therapeutic effect of taurine against aluminum-induced impairment on learning, memory and brain neurotransmitters in rats. *Neurol Sci*. 2014;35(10):1579-84. doi: 10.1007/s10072-014-1801-x, PMID 24770980.
- Ogunlade B, Adelakun SA, Agie JA. Nutritional supplementation of gallic acid ameliorates Alzheimer-type hippocampal neurodegeneration and cognitive impairment induced by aluminum chloride exposure in adult Wistar rats. *Drug Chem Toxicol*. 2022 Jun;45(2):651-62. doi: 10.1080/01480545.2020.1754849, PMID 32329360.
- Iqbal G, Iqbal A, Mahboob A, Farhat SM, Ahmed T. Memory enhancing effect of black pepper in the A β 1-42 induced neurotoxicity mouse model is mediated through its active component chavicine. *Curr Pharm Biotechnol*. 2016;17(11):962-73. doi: 10.2174/138920170666160709202124, PMID 27396401.
- Justin Thenmozhi A, Dhivyabharathi M, William Raja TR, Manivasagam T, Essa MM. Tannoid principles of *Emblica officinalis* renovate cognitive deficits and attenuate amyloid pathologies against aluminum chloride induced rat model of Alzheimer's disease. *Nutr Neurosci*. 2016;19(6):269-78. doi: 10.1179/1476830515Y.0000000016, PMID 25842984.
- Kumar V, Gill KD. Oxidative stress and mitochondrial dysfunction in aluminum neurotoxicity and its amelioration: a review. *Neurotoxicology*. 2014;41:154-66. doi: 10.1016/j.neuro.2014.02.004, PMID 24560992.
- Elshamy S, Abdel Motaal A, Abdel-Halim M, Medhat D, Handoussa H. Potential neuroprotective activity of *Mentha longifolia* L. in aluminum chloride-induced rat model of Alzheimer's disease. *J Food Biochem*. 2021;45(4):1770. doi: 10.1111/jfbc.13644, PMID 33587299.
- Busch CJ, Binder CJ. Malondialdehyde epitopes as mediators of sterile inflammation. *Biochim Biophys Acta Mol Cell Biol Lipids*. 2017;1862(4):398-406. doi: 10.1016/j.bbalip.2016.06.016, PMID 27355566.
- Mathiyazahan DB, Justin Thenmozhi A, Manivasagam T. Protective effect of black tea extract against aluminium chloride-induced Alzheimer's disease in rats: A behavioural, biochemical and molecular approach. *J Funct Foods*. 2015;16:423-35. doi: 10.1016/j.jff.2015.05.001.
- Abu-Elfotuh K, Ragab GM, Salahuddin A, Jamil L, Abd Al Haleem EN. Attenuative effects of fluoxetine and *Triticum aestivum* against aluminum-induced Alzheimer's disease in rats: the possible consequences on hepatotoxicity and nephrotoxicity. *Molecules*. 2021;26(21):6752. doi: 10.3390/molecules26216752, PMID 34771159.

39. Alghamdi BSA. Possible prophylactic anti-excitotoxic and anti-oxidant effects of virgin coconut oil on aluminium chloride-induced Alzheimer's in rat models. *J Integr Neurosci*. 2018;17(3-4):593-607. doi: 10.3233/JIN-180089, PMID 30010139.
40. Lin WT, Chen RC, Lu WW, Liu SH, Yang FY. Protective effects of low-intensity pulsed ultrasound on aluminum-induced cerebral damage in Alzheimer's disease rat model [sci rep]. *Sci Rep*. 2015;5(1):9671. doi: 10.1038/srep09671, PMID 25873429.
41. Justin Thenmozhi A, Raja TRW, Janakiraman U, Manivasagam T. Neuroprotective effect of hesperidin on aluminium chloride induced Alzheimer's disease in Wistar rats. *Neurochem Res*. 2015;40(4):767-76. doi: 10.1007/s11064-015-1525-1, PMID 25630717.
42. Satheeshkumar N, Mukherjee PK, Bhadra S, Saha BP. Acetylcholinesterase enzyme inhibitory potential of standardized extract of *Trigonella foenum Graecum* L and its constituents. *Phytomedicine*. 2010;17(3-4):292-5. doi: 10.1016/j.phymed.2009.06.006, PMID 19576740.

Cite this article: Li D, Man X, Ma X, Sun J. Neuroprotective Role of Periplocin Against Aluminium Chloride-stimulated Alzheimer's Disease in a Rat Model by Modulation of Oxidative Stress and Inflammation. *Ind. J. Pharm. Edu. Res.* 2023;57(1):147-54.