# Apocynin Induces Apoptosis in Human Lung Cancer A549 Cells via Regulating Apoptotic Protein and Inflammatory Cytokines

Li Shang<sup>1,#</sup>, Jindao Wang<sup>2,#</sup>, Xiaoqing Liu<sup>3,\*</sup>

- Department of Pulmonary and Critical Care Medicine, The People's Hospital of SND, Suzhou, Jiangsu, CHINA.
- <sup>2</sup>Department of Pulmonary and Critical Care Medicine, Huaian Hospital of Huaian City, Huaian Cancer Hospital, Huaian, Jiangsu, CHINA.
- <sup>3</sup>Department of Tumor Rehabilitation, Shaanxi Kangfu Hospital, Xi'an, Shaanxi, CHINA.
- \*The Authors Contributed Equally.

#### **ABSTRACT**

Objectives: Lung cancer, widely considered the leading cause of cancer-related deaths world wide, ranks fourth in terms of its occurrence among all types of malignant tumors. Phytochemicals are the potent alternative for these chemotherapeutic drugs. They have proven to render anticancer effects against various cancers such as breast, liver, brain, lung, skin, bone etc., in vivo, in vitro and even in clinical phase trials. One such phytochemical is apocynin, polyphenolic compound obtained from the Picrorhiza kurroa roots. It is a persuasive anti-inflammatory drug utilized to treat inflammatory ailments like arthritis, inflammatory bowel disease, colitis, stroke etc., Materials and Methods: We demonstrated the anticancer activity of apocynin against lung cancer A549 cells. Cytotoxicity effect of apocynin at different dose against A549 cells were assessed with MTT assay. The ROS accumulation and impairment of mitochondrial membrane permeability by apocynin in A549 cells were analyzed with DCFH-DA and Rhodamine 123 staining techniques. Dual staining with AO/EtBr was done to detect the percentage of cell apoptosis induced by apocynin in A549 cells. DAPI staining was performed to detect the induction of nuclear fragmentation in A549. The apoptosis induction and inflammation in A549 cells by apocynin was studied by quantifying the apoptotic proteins and inflammatory cytokines using ELISA technique. Results: Apocynin significantly induced cytotoxicity against A549 cells in dose dependent. It enhanced the caspases, Bax levels and diminished the antiapoptotic protein Bcl2 level. Apocynin also decreased the proinflammatory cytokines thereby prevented the cancer cell growth and aided apoptosis induction in A549. The induction of apoptosis in A549 cells were confirmed with our staining techniques. Conclusion: Our findings of apocynin on A549 cells suggest apocynin may possess possible therapeutic efficacy in treating lung cancer patients.

Keywords: Lung cancer, Phytochemical, Apocynin, A549 cells, Apoptosis, Inflammation.

### **Correspondence:**

#### Dr. Xiaoqing Liu

Department of Tumor Rehabilitation, Shaanxi Kangfu Hospital, Xi'an, Shaanxi-710065, CHINA. Email: liuxq92056302@outlook.com

**Received:** 06-06-2024; **Revised:** 03-07-2024; **Accepted:** 23-08-2024.

## **INTRODUCTION**

Lung cancer is a common type of cancer that arises from the mucous membrane of the bronchi, glands, or alveolar epithelium.¹ Lung cancer, widely considered the leading cause of cancer-related fatalities globally, ranks fourth in terms of its occurrence among all types of malignant tumors. In 2020, lung cancer was the leading cause of mortality among all types of cancer.² There are two primary forms of lung cancer: small cell lung cancer, which is less prevalent and Non-Small Cell Lung Cancer (NSCLC), which is responsible for the majority of lung cancer occurrences.³



**DOI:** 10.5530/ijper.20250045

#### Copyright Information :

Copyright Author (s) 2025 Distributed under Creative Commons CC-BY 4.0

Publishing Partner: Manuscript Technomedia.[www.mstechnomedia.com]

The incidence of lung cancer death worldwide is rapidly increasing due to the persistent adoption of cancer-causing behaviors such as smoking, physical inactivity and westernized diets. Lung cancer is the primary cause of cancer-related deaths in developing countries, surpassing other types of cancer such as colorectal, breast and prostate cancers.4 Lung cancer cells exhibit fast growth and are susceptible to distant metastases. Conventional cancer treatments, including surgical techniques, radiation and chemotherapy, have been utilized to treat lung cancer. However, the current traditional methods have proven ineffective in prolonging the survival of cancer patients.<sup>5</sup> The chemotherapeutic drugs have serious impacts causing various allergic reactions, neuropathic syndromes and hand-foot syndrome. Prolonged usage of chemotherapeutic drugs also leads to drug resistance in lung cancer patients impairing the drug potency. Furthermore, there have been significant advancements in the treatment of lung cancer, the prognosis for patients remains poor, since the radiotherapy associated with less than 20% of 5-year survival rate.<sup>6</sup> Therefore discovering a drug with high potency and minimal side effect is the prime goal for pharmaceutical industries to treat lung cancer.

Polyphenolic compounds are the predominant class of phytochemicals with proven pharmaceutical properties. These polyphenolic compounds possess high specificity and minimal toxicity hence it is under research to treat various diseases. They are potent antioxidant and anti-inflammatory agents are tending to be safe to treat neurological diseases. These compounds possess the property to alter the cancer cells epigenome thereby blocked cell cycle and triggered apoptosis in tumor cells. It also alters the antioxidant enzymes, apoptotic signaling protein in tumor cells inhibiting the proliferation growth and invasion of tumor cells. 100 per property to the proliferation growth and invasion of tumor cells. 100 per property to the proliferation growth and invasion of tumor cells. 100 per property to the proliferation growth and invasion of tumor cells. 100 per property to the proliferation growth and invasion of tumor cells. 100 per property to the pro

Apocynin is one such phenolic compound extracted from the *Apocynum cannabinum* and *Picrorhiza kurroa* roots which is a well-known inhibitor of NADPH-oxidase.<sup>11</sup> Numerous studies have proven the efficacy of apocynin on treating inflammatory disease such rheumatoid arthritis,<sup>12</sup> stroke,<sup>13</sup> atherosclerosis,<sup>14</sup> traumatic brain injury. It also shown promising effect on phase I clinical trial in treating anti-inflammatory diseases.<sup>15</sup> The anticancer efficacy of apocynin on lung cancer was not yet elucidated. In this work we assessed the potency of apocynin on human lung cancer cells.

## **MATERIALS AND METHODS**

### **Chemicals and Reagents**

Apocynin, with >98% of purity was procured from Sigma Aldrich USA. Supplements for culturing cells i.e., Dulbecco Modified Eagle Medium (DMEM), Fetal Bovine Serum (FBS), Dimethyl Sulfoxide (DMSO), TriZol reagent and antibiotic-antimycotic solution were obtained from ThermoFisher Scientific USA.

#### **Cell culture and Maintenance**

Lung cancer cells (A549) were viewed under microscope to examine the confluency of cells and any other contamination. After confirming the cells are free of contamination, the medium was changed and incubated in the 5% CO<sub>2</sub> supplemented incubator at 37°C. The cells were cultivated with DMEM supplemented with 10% FBS and 1% antimycotic solution. The cells were trypsinized using Trypsin solution for further sub culturing.

## **Cell cytotoxicity Assay**

A549 cells were trypsinized with trypsin-EDTA and  $1x10^4$  cells/mL were loaded on to 96-wellplate and incubated for 24 hr. The cultured cells were treated with diverse dosages of apocynin ranging from 10-100  $\mu$ M/mL in triplicates and incubated for 24 hr at 37°C in the 5%  $CO_2$  supplemented incubator. After 24 hr the cells were investigated by MTT

(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. The 20  $\mu$ L of MTT reagent with 5 mg/mL concentration was mixed to the cultured cells and incubated at 37°C in the 5% CO $_2$  supplemented incubator for 4 hr. After incubation period the culture medium was discarded and the formazan deposits was liquefied with 0.1 mL/well of DMSO solution. The final absorbance was taken at 490 nm. The experiment was repeated thrice and % of viability was determined by following equation

Viability (%)=OD of treated cells/OD of control cellsX100

## **Quantification of Reactive Oxygen Species (ROS)**

ROS generated by the apocynin treated and control A549 cells were quantified using 2,7-Dichlorodihydrofluorescein Diacetate (DCFH-DA) staining technique (Invitrogen, USA). The A549 cells were exposed to 40 and 60  $\mu$ M/mL dosages of apocynin for 24 hr at 37°C in 5% CO $_2$  supplemented incubator. After incubation the cells were treated with 10 mM DCFH-DA dye for 30 min. The stained cells were rinsed with Phosphate Buffered Saline (PBS) and monitored using fluorescent microscope. The images were then analyzed with ImageJ software and the proportion of fluorescence intensity was determined.

# **Quantification of Mitochondrial Membrane Potential** (MMP)

The MMP of control and apocynin treated A549 cells was investigated using Rhodamine-123 stain (Invitrogen, USA). The control, 40 and 60  $\mu\text{M/mL}$  apocynin treated A549 cells were Rhodamine-123 dye for 30 min. Later, the cells were rinsed twice with PBS and observed using fluorescent microscope. The images were further assessed with ImageJ software to quantify the intensity of fluorescence.

## **Quantification of Live/apoptotic cells**

The apoptosis induction by apocynin in human lung cancer A549 cells was measured using dual staining technique. A549 cells were exposed with 40 and 60  $\mu M/mL$  of apocynin for 24 hr at 37°C. The cells were then treated with 1  $\mu g/mL$  of AO and EtBr (Sigma Aldrich, USA) in 1:1 ratio and incubated for 30 min. The cell was then washed with PBS and scrutinized using fluorescence microscope. The percentage of live and dead cells was measured using ImageJ software.

## **Quantification of nuclear damage**

The nuclear damage induced by apocynin on A549 cells were quantified using the nuclear stain DAPI (Invitrogen, USA). The 40 and 60  $\mu\text{M/mL}$  of apocynin treated cell were stained with nuclear counterstain 300 nM DAPI and incubated for 15 min in dark. The stain was removed after incubation and rinsed with PBS thrice. The cells were then investigated using fluorescence microscope and percentage of apoptotic cells were quantified using ImageJ software.

243

## **Quantification of apoptotic protein**

The Caspase-3, Caspase-9, Bax and Bcl2 expressions were quantified using ELISA kits procured from Abcam, USA. Human lung cancer A549 cells were exposed to 40 and 60 µM dosages of apocynin for 24 hr at 37°C in 5% CO<sub>2</sub> supplemented incubator. Later, the cells were trypsinized and subjected to sonication. The sonicated cells were centrifuged at 10000 rpm for 10 min at 4°C. Supernatant was collected for further quantification of apoptotic proteins. The reagents and the standards were prepared freshly 30 min before the experiment. The ELISA plates were washed twice with the saline before the initiation of experiment. The 100 µL of sample was mixed to the wells in triplicates and incubated at 37°C for 45 min. After incubation the wells were emptied gently by pipetting and add 100 µL of biotin-coated antibody work solution. The plates were covered and incubated for 1 hr at 37°C. The wells were discarded after incubation and washed thrice with 1X wash buffer. The 100 µL of SABC working solution was mixed to the decanted wells and incubated at 37°C for 30 min. The suspension was removed and wells were rinsed thrice with saline. To the wells  $90 \mu L$  of TMB substrate were mixed and incubated the plates in dark at 37°C for 20 min. Finally, 50 μL of stop solution was mixed to the wells and absorbance was taken at 450 nm. Standard curve was plotted with the OD 450 values of standards in Y axis Vs the respective concentration in X axis. The concentrations of samples were measured using the standard curve.

## **Quantification of inflammatory cytokines**

The inflammatory cytokine levels TNF- $\alpha$ , NF $\kappa$ B, COX-2 and IL-6 were quantified in the control and apocynin treated A549 cells using the ELISA kits procured from Abcam, USA. Same protocol mentioned for quantification of apoptotic proteins were carried for the quantification of inflammatory cytokines also.

### **Statistical Analysis**

The studies were performed thrice and values obtained was scrutinized with the statistical software GraphPad Prism, version 6.0. The intergroup difference was investigated with one way ANOVA and the intragroup significance was assessed with *post hoc* Tukey's test. The significance was set to be p<0.05 and the data were presented as mean±SD.

### **RESULTS**

# Polyphenol apocynin induced cytotoxicity in A549 cells

Figure 1 illustrates the results obtained from the MTT assay performed with untreated and apocynin treated A549 cells. Apocynin considerably inhibited the viability of A549 in a dose-dependent manner. Control untreated A549 cells were considered as 100% viable and the percentage of cell viability in apocynin treated were calculated accordingly. Minimal dose of 10  $\mu$ M/mL of apocynin treated cells shown 97±2.2% of cell

viability whereas the highest dose treated 100  $\mu$ M/mL apocynin treated cells shown minimum of 18±1.8% of cell viability. IC<sub>50</sub> level was fixed at 40  $\mu$ M/mL of apocynin.

# Polyphenol apocynin induces ROS generation in A549 cells

The levels of reactive oxygen generated in untreated and apocynin treated A549 cells were measured by quantifying the green fluorescence emitted due to DCFH-DA staining. Apocynin treated A549 cells emitted increased level of fluorescence in doses dependent manner indicating the accumulation of ROS. The 40  $\mu$ M/mL apocynin treated cells shown 29±1.2% of fluorescence whereas it is significantly increased in to 52±2.3% of fluorescence in 60  $\mu$ M/mL apocynin treated A549 cells (Figure 2).

## Polyphenol apocynin decrease MMP in A549 cells

Mitochondrial activity depends on the mitochondrial membrane potential hence we assessed the role of apocynin on disrupting the MMP potential in A549 cells. Control untreated cells fluorescence was considered to be 100%. The 40  $\mu$ M/mL apocynin treated cells shown 69±0.8% of fluorescence whereas it is significantly decreased to 53±0.9% of fluorescence in 60  $\mu$ M/mL apocynin treated A549 cells (Figure 3).

# Polyphenol apocynin induced apoptosis in A549 cells

Figure 4 depicts the representative AO/EtBr-stained images of A549 cells. Control cells shown bright green fluorescence indicating the presence of viable cells. The 40  $\mu$ M/mL apocynin treated cells displayed greenish orange fluorescence and the 60  $\mu$ M/mL of apocynin treated cells exhibited bright red orange fluorescence indicating the increased number of apoptotic cells. The 73±1.2% of viable and 25±0.9% of dead cells were detected in 40  $\mu$ M/mL apocynin treated cells whereas the 60  $\mu$ M/mL apocynin treatment significantly decreased the viable cell percentage to 30±1.4.

# Polyphenol apocynin induced nuclear damage in A549 cells

Polyphenol apocynin treated A549 cells were subjected to staining with nuclear stain DAPI to assess the nuclear damage induced by apocynin (Figure 5). Both the 40 and 60  $\mu M$  apocynin treated A549 cells exhibited upsurge in number of fragmented nucleuses containing cells that proves the induction of apoptosis. The results of ImageJ analysis confirms that 38±0.7% and 57±0.9% of cells were apoptotic cells in 40 and 60  $\mu M/mL$  apocynin treated cells, respectively.

# Polyphenol apocynin induced apoptosis in A549 cells

The apoptosis induction by apocynin in A549 cells were analyzed by estimating the apoptotic proteins Caspase-9, Caspase-3, Bax

and Bcl2 levels using ELISA kits (Figure 6). Both the initiator caspase-9 and executor caspase-3 were significantly augmented in the apocynin treated cells than untreated lung cancer cells. The proapoptotic protein Bax levels were also increased in the apocynin treated A549 cells. The antiapoptotic protein Bcl2 levels were significantly decreased in the apocynin exposed cells than control cells.

## Polyphenol apocynin induced inflammation in A549 cells

Inflammation plays a crucial role in tumor development hence we assessed the levels of inflammatory protein in apocynin treated human lung cancer cells (Figure 7). Apocynin treatment remarkably diminished the levels of proinflammatory cytokines TNF $\alpha$  and NF $\kappa$ B. It also decreased the interleukin-6 and cyclooxygenase levels that confirming the anti-inflammatory potency of apocynin.

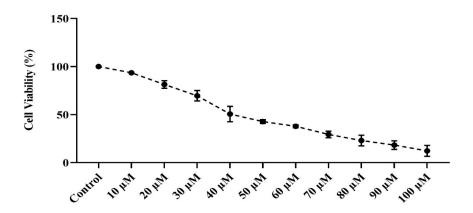


Figure 1: Polyphenol apocynin induced cytotoxicity in human lung cancer A549 cells. A549 cells were cultured in DMEM medium and treated with different concentrations of apocynin 10-100  $\mu$ M/mL for 24 hr. Subjected to MTT assay in triplicates and the results were assessed with one way ANOVA followed by *post hoc* Tukey's test. Data were presented as mean±SD, p<0.05 considered to be statistically significant.

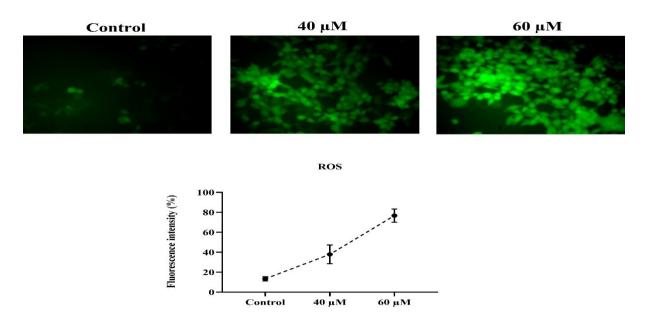


Figure 2: Polyphenol apocynin induces ROS generation in human lung cancer A549 cells. A549 cells were cultured in DMEM medium and treated with two different concentrations of 40 and 60  $\mu$ M/mL apocynin for 24 hr. Apocynin treated cells were subjected to 2,7-Dichlorodihydrofluorescein Diactate (DCFH-DA) staining. The images were assessed with ImageJ software. Experiments were done in triplicates and the results were assessed with one way ANOVA followed by *post hoc* Tukey's test. Data were presented as mean $\pm$ SD, p<0.05 considered to be statistically significant.

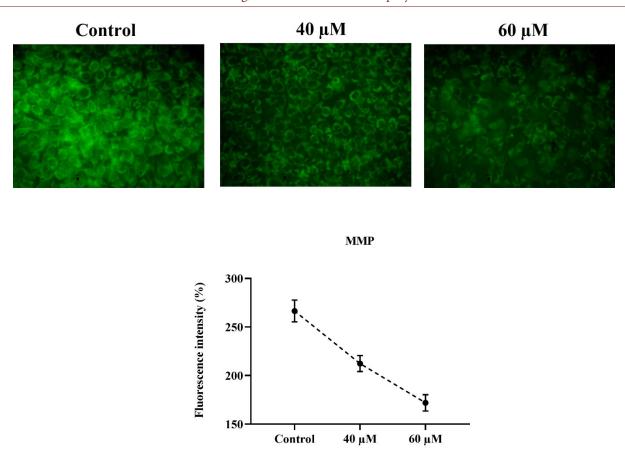


Figure 3: Polyphenol apocynin decrease mitochondria membrane potential in human lung cancer A549 cells. A549 cells were cultured in DMEM medium and treated with two different concentrations of 40 and 60 μM/mL apocynin for 24 hr. Apocynin treated cells were subjected to Rhodamine 123 staining. The images were assessed with ImageJ software. Experiments were done in triplicates and the results were assessed with one way ANOVA followed by *post hoc* Tukey's test. Data were presented as mean±SD, *p*<0.05 considered to be statistically significant

#### DISCUSSION

In the current study, we attempted to analyze the potency of the phytochemical apocynin to induce apoptosis in lung cancer cells. Lung cancer is one of the deadliest diseases which were ranked to second in cancer related global mortalities lack efficient treatment. The major drawback of treating lung cancer is lack of diagnosis at early stages.<sup>3</sup> Most of lung cancer patients were diagnosed at the metastasis stage in which the traditional treatments such as surgery, radiotherapy and chemotherapy are in effective.<sup>4</sup> Even though some chemotherapeutic drugs are available to treat lung cancer it induces numerous side effects which is worse than the cancer induced health morbidities.

Apocynin is polyphenolic compound isolated from *Picrorhiza kurroa* perennial plant present in Himalayas.<sup>16</sup> The extracts of this medicinal plant were used in the ancient ayurvedic medicine to treat diseases related to liver, lung, heart and joints. Apocynin is effective Nicotinamide Adenine Dinucleotide Phosphate (NADPH)-oxidase inhibitor.<sup>17-20</sup> Therefore it is proven to be potent against inflammatory diseases such arthritis, neurodegenerative diseases, stroke, asthma, inflammatory bowel disease, Crohn

disease, colitis etc.,<sup>21</sup> Few studies have reported apocynin inhibits the cancer cell progression in prostate,<sup>22</sup> breast<sup>23</sup> and pancreatic cancer cells.<sup>24</sup> We analyzed the role apocynin against lung cancer cells.

The impact of apocynin on cell survival of A549 cells were examined by treating the cells with different concentration of apocynin for 24 hr and subjected to MTT assay. At 24 hr incubation apocynin effectively inhibited the survival of lung cancer and induced 50% of cell death at the dose of 40  $\mu$ M/mL concentration. Hence for assessment of the anticancer property we treated the A549 cells with a dose of 40 and 60  $\mu$ M/mL which is lesser than the IC<sub>50</sub> dose of apocynin on A549 cells.

ROS plays a vital role in cellular homeostasis maintenance and the dysregulation in ROS and antioxidants leads to cancer cell proliferation and progression. ROS is a double-edged sword not only induces the tumor cell development but in certain threshold it also triggers apoptosis in tumor cells. Hence most of the anticancer drugs are designed to enhance the ROS in cancer cells thereby inducing cancer cell apoptosis.<sup>25-28</sup> It often impairs the antioxidant system in cancer cells which aids the drug resistance

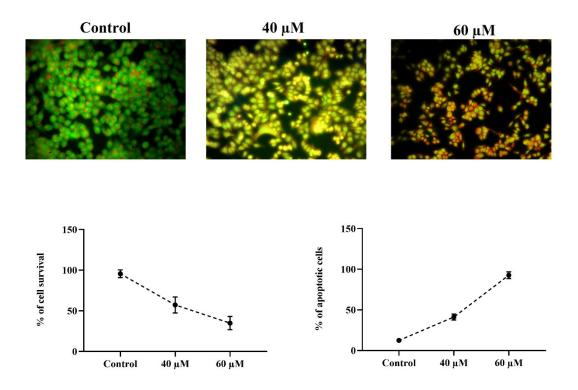


Figure 4: Polyphenol apocynin induced cell death in human lung cancer A549 cells. A549 cells were cultured in DMEM medium and treated with two different concentrations of 40 and 60  $\mu$ M/mL apocynin for 24 hr. Apocynin treated cells were subjected to dual staining with acridine orange and ethidium bromide mixture (1:1). The images were assessed with ImageJ software. Experiments were done in triplicates and the results were assessed with one way ANOVA followed by *post hoc* Tukey's test. Data were presented as mean±SD, p<0.05 considered to be statistically significant.

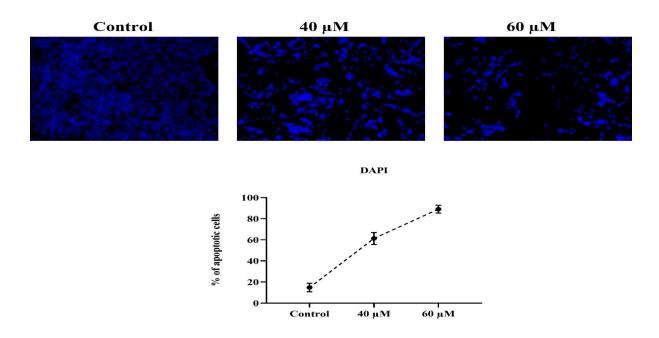


Figure 5: Polyphenol apocynin induced nuclear damage in human lung cancer A549 cells. A549 cells were cultured in DMEM medium and treated with two different concentrations of 40 and 60 μM/mL apocynin for 24 hr. Apocynin treated cells were subjected to DAPI staining. The images were assessed with ImageJ software. Experiments were done in triplicates and the results were assessed with one way ANOVA followed by post hoc Tukey's test. Data were presented as mean±SD, p<0.05 considered to be statistically significant.

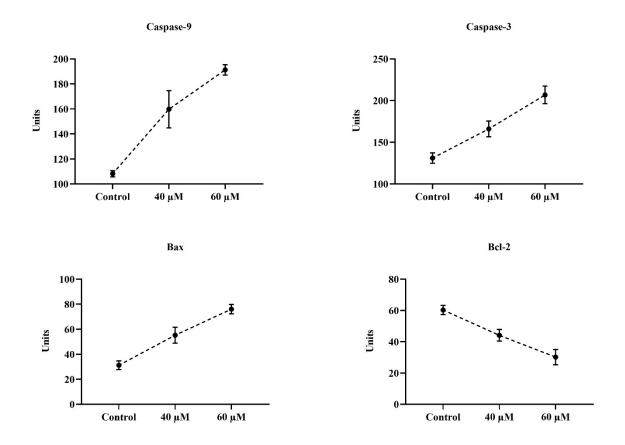


Figure 6: Polyphenol apocynin induced apotosis in human lung cancer A549 cells. A) Caspase 9, B) Caspase 3 C) Bax D) Bcl2. A549 cells were cultured in DMEM medium and treated with two different concentrations of 40 and 60 μM/mL apocynin for 24 hr. Apocynin treated cells were subjected to quantification of apoptotic proteins using ELISA kits. Experiments were done in triplicates and the results were assessed with one way ANOVA followed by *post hoc* Tukey's test. Data were presented as mean±SD, *p*<0.05 considered to be statistically significant.

in cancer cells. The presence of increased antioxidants like GSH as glutathione impairs the drug potency of anticancer drugs such taxol. Arsenic trioxide etc., Anticancer drug Procarbazine induces apoptosis in Hodgkin's lymphoma via ROS induced oxidative nuclear damage.<sup>29</sup> Doxorubicin a potent anticancer prescribed to treat various cancers such as breast, Kaposi's sarcoma, leukemia, bladder stimulates the Fenton reaction thereby induces the generation of hydroxyl radicals causing oxidative stress induced apoptosis.<sup>30,31</sup> Hence we assessed the efficacy of apocynin to induce ROS generation in A549 cells. Apocynin significantly increased the levels of ROS in A549 cells.

Increase in ROS acts on sphingomyelinase induces the synthesis of ceramide which binds to the cell membrane death receptors of cancer cells causing apoptosis. Lymphoma cells induced with UV radiation activates this pathway thereby collapses mitochondria and leads to apoptosis.<sup>32</sup> Drugs such as gamitrinib induces mitochondrial damage in prostate<sup>33</sup> while quinone derivative drug ARQ501 disrupts electron transport chain thereby leads to apoptosis in pancreatic cancers.<sup>34</sup> Apocynin treatment also disrupted the MMP in A549 cells which was evidenced by Rhodamine 123 staining results. The disruption of MMP

may because of the increase in ROS induced by the apocynin treatment.

Phytochemicals like resveratrol, thymoquinone, ginsenosides acts as prooxidants induces generation and generation of ROS in mitochondria of cancer cells such as PC-3, MCF-7, HepG2 thereby induces apoptosis.35,36 ROS triggers various signaling pathways activates Akt which further activates apoptotic protein Bax. Activated Bax disrupts the MMP and release cytochrome-C into the cytosol.<sup>37</sup> In cytosol cytochrome activates procaspase -9 and forms apoptosomes. It in turn activates effector caspase 3 which cleaves the cellular protein causing apoptosis. 38 In our study apocynin treatment significantly increased Bax and also both the initiator caspase-9 and the effector Caspase 3 which confirms the activation of apoptotic signaling pathway. The activation of these apoptotic proteins may be due to the increased ROS accumulation which was confirmed with DCFH-DA staining results. Further the results of DAPI and AO/EtBr staining proves the initiation of apoptosis in A549 cells by apocynin.

Inflammation is another major entity which regulates the cancer cell growth and progression. The levels of inflammatory cytokines in the tumor cells determines the transformation of

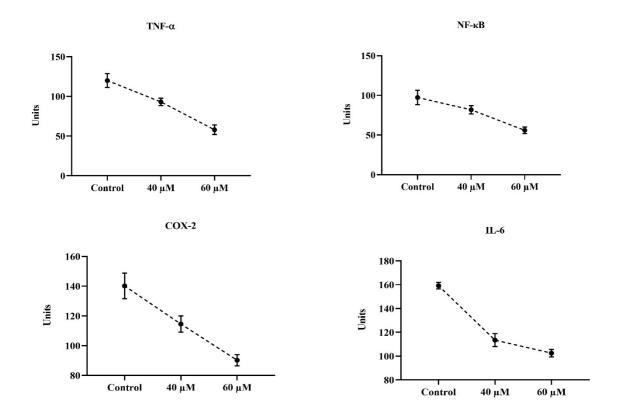


Figure 7: Polyphenol apocynin induced inflammation in human lung cancer A549 cells. A) Tumor necrosis factor alpha-TNF $\alpha$  B) Nuclear factor kappa B-NF $\alpha$ B C) Cyclooxygenase -2 COX-2 D) Interleukin-6 IL-6. A549 cells were cultured in DMEM medium and treated with two different concentrations of 40 and 60 μM/mL apocynin for 24 hr. Apocynin treated cells were subjected to quantification of inflammatory cytokines using ELISA kits. Experiments were done in triplicates and the results were assessed with one way ANOVA followed by *post hoc* Tukey's test. Data were presented as mean $\pm$ SD, p<0.05 considered to be statistically significant.

malignant cells and also induces antitumor response.<sup>39-41</sup> ROS is key intermediator between the inflammatory cytokines and cancer cells it regulates the levels and activation of inflammatory cytokines. 42,43 ROS accumulation, Chronic inflammation and cancer cell progression are interlinked signaling pathways. ROS promotes TNF-α and NFκB levels, the activated NFκB is the prime inducer of drug resistance for various mostly prescribed anticancer drugs such as tamoxifen, cisplatin, daunorubicin, 5-fluorouracil etc., The activation of NF-κB up regulates cell survival proteins COX-2, Bcl2, Akt, cyclin D1, surviving. 44-46 Therefore anticancer drugs were targeted to increase the ROS production and decrease the inflammatory cytokines thereby inhibit the cancer proliferation and metastasis. 47,48 In our work also apocynin remarkably augmented the ROS accumulation and decreased the TNF-α, NFκB, COX-2 and IL-6 that confirms it may be a talented anticancer candidate to treat human lung cancer.

## **CONCLUSION**

To conclude our findings suggests phytochemical apocynin significantly increased the ROS generation in A549 cells which in turn disrupted the mitochondrial membrane potential and

triggered the apoptosis signaling molecules. It also effectively diminished the proinflammatory cytokines in A549. Apocynin is a talented anticancer and anti-inflammatory drug which possess the potency to be developed as anticancer agent to treat human lung cancer in future.

### **ACKNOWLEDGEMENT**

This work was supported by the Department of Tumor Rehabilitation, Shaanxi Kangfu Hospital, Xi'an, Shaanxi, 710065, China.

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

## **ABBREVIATIONS**

NSCLC: Non-small cell lung cancer; DMEM: Dulbecco's Modified Eagle's Medium; FBS: Fetal bovine serum; DMSO: Dimethylsulfoxide; DCFH-DA: 2,7-dichlorodihydrofluorescein diactate; MMP: Mitochondrial Membrane Potential.

### **SUMMARY**

Apocynin is one such phenolic compound extracted from the *Apocynum cannabinum* and *Picrorhiza kurroa* roots which are numerous studies have proven the efficacy of apocynin on treating various disease. Lung cancer, widely considered the leading cause of cancer-related fatalities globally, ranks fourth in terms of its occurrence among all types of malignant tumors. apocynin significantly increased the ROS generation in A549 cells which in turn disrupted the mitochondrial membrane potential and triggered the apoptosis signaling molecule. Apocynin remarkably augmented the ROS accumulation and decreased the TNF- $\alpha$ , NFkB, COX-2 and IL-6 that confirms it may be a talented anticancer candidate to treat human lung cancer.

#### REFERENCES

- Bolgova LS, Tuganova TN, Alekseenko OI, Litvinets OM, Suprun GA, Ponomarenko AA. Histogenesis of central lung cancer: cytological investigation. Exp Oncol. 2020;42(4):310-3. doi: 10.32471/exp-oncology.2312-8852.vol-42-no-4.15232, PMID 33355875.
- Schabath MB, Cote ML. Cancer progress and priorities: lung cancer. Cancer Epidemiol Biomarkers Prev. 2019;28(10):1563-79. doi: 10.1158/1055-9965.EPI-19-0221, PMID 31575553
- Blandin Knight S, Crosbie PA, Balata H, Chudziak J, Hussell T, Dive C. Progress and prospects of early detection in lung cancer. Open Biol. 2017;7(9):170070. doi: 10.109 8/rsob.170070. PMID 28878044.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209-49. doi: 10.3322/ca ac.21660, PMID 33538338.
- 5. Barta JA, Powell CA, Wisnivesky JP. Global epidemiology of lung cancer. Ann Glob Health. 2019;85(1):8. doi: 10.5334/aogh.2419, PMID 30741509.
- Wadowska K, Bil-Lula I, Trembecki Ł, Śliwińska-Mossoń M. Genetic markers in lung cancer diagnosis: a review. Int J Mol Sci. 2020;21(13):4569. doi: 10.3390/ijms211345 69, PMID 32604993.
- Naseri R, Farzaei F, Fakhri S, El-Senduny FF, Altouhamy M, Bahramsoltani R, et al. Polyphenols for diabetes associated neuropathy: pharmacological targets and clinical perspective. Daru. 2019;27(2):781-98. doi: 10.1007/s40199-019-00289-w, PMID 31352568.
- Abbaszadeh F, Fakhri S, Khan H. Targeting apoptosis and autophagy following spinal cord injury: therapeutic approaches to polyphenols and candidate phytochemicals. Pharmacol Res. 2020;160:105069. doi: 10.1016/j.phrs.2020.105069, PMID 32652198.
- Gavrilas LI, Cruceriu D, Mocan A, Loghin F, Miere D, Balacescu O. Plant-derived bioactive compounds in colorectal cancer: insights from combined regimens with conventional chemotherapy to overcome drug-resistance. Biomedicines. 2022;10(8):1948. doi: 10.3390/biomedicines10081948, PMID 36009495.
- Sharma A, Kaur M, Katnoria JK, Nagpal AK. Polyphenols in food: cancer prevention and apoptosis induction. Curr Med Chem. 2018;25(36):4740-57. doi: 10.2174/092986 7324666171006144208, PMID 28990504.
- Nasir A, Bullo MM, Ahmed Z, Imtiaz A, Yaqoob E, Jadoon M, et al. Nutrigenomics: epigenetics and cancer prevention: A comprehensive review. Crit Rev Food Sci Nutr. 2020;60(8):1375-87. doi: 10.1080/10408398.2019.1571480, PMID 30729798.
- White CN, Figtree GA, Liu CC, Garcia A, Hamilton EJ, Chia KK, et al. Angiotensin II inhibits the Na+-K+ pump via PKC-dependent activation of NADPH oxidase. Am J Physiol Cell Physiol. 2009; 296(4):C693-700. doi: 10.1152/ajpcell.00648.2008, PMID 19193863.
- 13. Kryl'skii ED, Popova TN, Kirilova EM. Activity of glutathione antioxidant system and NADPH-generating enzymes in rats with experimental rheumatoid arthritis. Bull Exp Biol Med. 2015;160(1):24-7. doi: 10.1007/s10517-015-3089-0, PMID 26601844.
- Choi BY, Jang BG, Kim JH, Lee BE, Sohn M, Song HK, et al. Prevention of traumatic brain injury-induced neuronal death by inhibition of NADPH oxidase activation. Brain Res. 2012; 1481:49-58. doi: 10.1016/j.brainres.2012.08.032, PMID 22975130.
- Kinkade K, Streeter J, Miller FJ. Inhibition of NADPH oxidase by apocynin attenuates progression of atherosclerosis. Int J Mol Sci. 2013;14(8):17017-28. doi: 10.3390/ijms1 40817017, PMID 23965970.
- Sun Y, Gong F, Yin J, Wang X, Wang X, Sun Q, et al. Therapeutic effect of apocynin through antioxidant activity and suppression of apoptosis and inflammation after spinal cord injury. Exp Ther Med. 2017;13(3):952-60. doi: 10.3892/etm.2017.4090, PMID 28450925.

- 't Hart BA, Copray S, Philippens I. Apocynin, a low molecular oral treatment for neurodegenerative disease. BioMed Res Int. 2014; 2014;298020. doi: 10.1155/2014 /298020. PMID 25140304.
- Qin YY, Li M, Feng X, Wang J, Cao L, Shen XK, et al. Combined NADPH and the NOX inhibitor apocynin provides greater anti-inflammatory and neuroprotective effects in a mouse model of stroke. Free Radic Biol Med. 2017;104:333-45. doi: 10.1016/j.free radbiomed.2017.01.034, PMID 28132925.
- Min S, Kim OJ, Bae J, Chung TN. Effect of pretreatment with the NADPH oxidase inhibitor apocynin on the therapeutic efficacy of human placenta-derived mesenchymal stem cells in intracerebral hemorrhage. Int J Mol Sci. 2018;19(11):3679. doi: 10.3390/ijms19113679, PMID 30469327.
- Du ZD, Yu S, Qi Y, Qu TF, He L, Wei W, et al. NADPH oxidase inhibitor apocynin decreases mitochondrial dysfunction and apoptosis in the ventral cochlear nucleus of D-galactose-induced aging model in rats. Neurochem Int. 2019;124:31-40. doi: 10 .1016/j.neuint.2018.12.008, PMID 30578839.
- Hou L, Sun F, Huang R, Sun W, Zhang D, Wang Q. Inhibition of NADPH oxidase by apocynin prevents learning and memory deficits in a mouse Parkinson's disease model. Redox Biol. 2019;22:101134. doi: 10.1016/j.redox.2019.101134, PMID 30798073.
- Stefanska J, Pawliczak R. Apocynin: molecular aptitudes. Mediators Inflamm. 2008; 2008:106507. doi: 10.1155/2008/106507, PMID 19096513.
- Suzuki S, Pitchakarn P, Sato S, Shirai T, Takahashi S. Apocynin, an NADPH oxidase inhibitor, suppresses progression of prostate cancer via Rac1 dephosphorylation. Exp Toxicol Pathol. 2013;65(7-8):1035-41. doi: 10.1016/j.etp.2013.03.002, PMID 23664830.
- Klees RF, De Marco PC, Salasznyk RM, Ahuja D, Hogg M, Antoniotti S, et al. Apocynin derivatives interrupt intracellular signaling resulting in decreased migration in breast cancer cells. J Biomed Biotechnol. 2006;2006(2):87246. doi: 10.1155/JBB/2006/8724 6, PMID 16883056.
- Kato A, Naiki-Ito A, Nakazawa T, Hayashi K, Naitoh I, Miyabe K, et al. Chemopreventive effect of resveratrol and apocynin on pancreatic carcinogenesis via modulation of nuclear phosphorylated GSK3β and ERK1/2. Oncotarget. 2015;6(40):42963-75. doi: 1 0.18632/oncotarget.5981, PMID 26556864.
- Pan JS, Hong MZ, Ren JL. Reactive oxygen species: a double-edged sword in oncogenesis. World J Gastroenterol. 2009;15(14):1702-7. doi: 10.3748/wjg.15.1702, PMID 19360913.
- Trachootham D, Alexandre J, Huang P. Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach? Nat Rev Drug Discov. 2009;8(7):579-91. doi: 10.1038/nrd2803. PMID 19478820.
- Tong L, Chuang CC, Wu S, Zuo L. Reactive oxygen species in redox cancer therapy. Cancer Lett. 2015;367(1):18-25. doi: 10.1016/j.canlet.2015.07.008, PMID 26187782.
- Yang H, Villani RM, Wang H, Simpson MJ, Roberts MS, Tang M, et al. The role of cellular reactive oxygen species in cancer chemotherapy. J Exp Clin Cancer Res. 2018;37(1):266. doi: 10.1186/s13046-018-0909-x. PMID 30382874.
- Renschler MF. The emerging role of reactive oxygen species in cancer therapy. Eur J Cancer. 2004;40(13):1934-40. doi: 10.1016/j.ejca.2004.02.031, PMID 15315800.
- 31. Kotamraju S, Chitambar CR, Kalivendi SV, Joseph J, Kalyanaraman B. Transferrin receptor-dependent iron uptake is responsible for doxorubicin-mediated apoptosis in endothelial cells: role of oxidant-induced iron signaling in apoptosis. J Biol Chem. 2002;277(19):17179-87. doi: 10.1074/jbc.M111604200, PMID 11856741.
- 32. Perillo B, Di Donato M, Pezone A, Di Zazzo E, Giovannelli P, Galasso G, et al. ROS in cancer therapy: the bright side of the moon. Exp Mol Med. 2020;52(2):192-203. doi: 1 0.1038/s12276-020-0384-2. PMID 32060354.
- Charruyer A, Grazide S, Bezombes C, Müller S, Laurent G, Jaffrézou JP. UV-C light induces raft-associated acid sphingomyelinase and JNK activation and translocation independently on a nuclear signal. J Biol Chem. 2005;280(19):19196-204. doi: 10.107 4/ibc.M412867200. PMID 15769735.
- Kang BH, Siegelin MD, Plescia J, Raskett CM, Garlick DS, Dohi T, et al. Preclinical characterization of mitochondria-targeted small molecule hsp90 inhibitors, gamitrinibs, in advanced prostate cancer. Clin Cancer Res. 2010;16(19):4779-88. doi: 10.1158/1078-0432.CCR-10-1818, PMID 20876793.
- Sborov DW, Haverkos BM, Harris PJ. Investigational cancer drugs targeting cell metabolism in clinical development. Expert Opin Investig Drugs. 2015;24(1):79-94. doi: 10.1517/13543784.2015.960077, PMID 25224845.
- Khan MA, Chen HC, Wan XX, Tania M, Xu AH, Chen FZ, et al. Regulatory effects of resveratrol on antioxidant enzymes: a mechanism of growth inhibition and apoptosis induction in cancer cells. Mol Cells. 2013;35(3):219-25. doi: 10.1007/s10059-013-2259-z, PMID 23456297.
- Sodrul IMD, Wang C, Chen X, Du J, Sun H. Role of ginsenosides in reactive oxygen species-mediated anticancer therapy. Oncotarget. 2017;9(2):2931-50.
- Sritharan S, Sivalingam N. Curcumin induced apoptosis is mediated through oxidative stress in mutated p53 and wild type p53 colon adenocarcinoma cell lines. J Biochem Mol Toxicol. 2021;35(1):22616. doi: 10.1002/jbt.22616, PMID 32864863.
- Redza-Dutordoir M, Averill-Bates DA. Activation of apoptosis signalling pathways by reactive oxygen species. Biochim Biophys Acta. 2016 Dec;1863(12):2977-92. doi: 10.1 016/j.bbamcr.2016.09.012, PMID 27646922.
- Gonda TA, Tu S, Wang TC. Chronic inflammation, the tumor microenvironment and carcinogenesis. Cell Cycle. 2009;8(13):2005-13. doi: 10.4161/cc.8.13.8985, PMID 19550141.

- Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and cytokines in the tumor microenvironment. J Immunol Res. 2014; 2014:149185. doi: 10.1155/2014/149185, PMID 24901008.
- 42. Singh R, Mishra MK, Aggarwal H. Inflammation, immunity and cancer. Mediators Inflamm. 2017; 2017:6027305. doi: 10.1155/2017/6027305, PMID 29234189.
- 43. Forrester SJ, Kikuchi DS, Hernandes MS, Xu Q, Griendling KK. Reactive oxygen species in metabolic and inflammatory signaling. Circ Res. 2018;122(6):877-902. doi: 10.1161 /CIRCRESAHA.117.311401, PMID 29700084.
- Aggarwal V, Tuli HS, Varol A, Thakral F, Yerer MB, Sak K, et al. Role of reactive oxygen species in cancer progression: molecular mechanisms and recent advancements. Biomolecules. 2019;9(11):735. doi: 10.3390/biom9110735, PMID 31766246.
- 45. Manna SK, Aggarwal RS, Sethi G, Aggarwal BB, Ramesh GT. Morin (3,5,7,2',4'-pentahydroxyflavone) abolishes nuclear factor-kappaB activation
- induced by various carcinogens and inflammatory stimuli, leading to suppression of nuclear factor-kappaB-regulated gene expression and up-regulation of apoptosis. Clin Cancer Res. 2007;13(7):2290-7. doi: 10.1158/1078-0432.CCR-06-2394, PMID 17404114
- 46. AhnKS, SethiG, Chaturvedi MM, Aggarwal BB. Simvastatin, 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitor, suppresses osteoclastogenesis induced by receptor activator of nuclear factor-kappaB ligand through modulation of NF-kappaB pathway. Int J Cancer. 2008;123(8):1733-40. doi: 10.1002/ijc.23745, PMID 18688862.
- 47. Zheng HC. The molecular mechanisms of chemoresistance in cancers. Oncotarget. 2017;8(35):59950-64. doi: 10.18632/oncotarget.19048, PMID 28938696.
- 48. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646-74. doi: 10.1016/j.cell.2011.02.013, PMID 21376230.

Cite this article: Shang L, Wang J, Liu X. Apocynin Induces Apoptosis in Human Lung Cancer A549 Cells via Regulating Apoptotic Protein and Inflammatory Cytokines. Indian J of Pharmaceutical Education and Research. 2025;59(1):242-51.