

Network Pharmacology-Based Modeling of Phytocompounds in a Traditional *Siddha* Formulation-*Kabasura kudineer* with Special Reference to SARS-CoV-2

Krishna Kant Gupta¹, Yatindrapravanan Narasimhan¹, Shashank Ravichandran¹, Brindha Pemaiah², Davidraj Chellappan², Ragothaman Madhava Rao Yennamalli^{1,*}, Sriram Sridharan^{2,*}

¹Department of Bioinformatics, School of Chemical and Biotechnology, SASTRA (Deemed to be University), Thanjavur, Tamil Nadu, INDIA.

²Centre for Advanced Research in Indian System of Medicine (CARISM), School of Chemical and Biotechnology, SASTRA (Deemed to be University), Thanjavur, Tamil Nadu, INDIA.

ABSTRACT

Background: *Kabasuk kudineer* (Kk), a traditional *Siddha* formulation containing 15 plant-based ingredients has been prescribed in *Siddha* medicine for the management and treatment of flu-like symptoms. Currently, the Ministry of AYUSH, Government of India has been prescribing Kk as a possible preventive and prophylactic formulation against COVID-19. **Objectives:** The present study focuses on computational methods and aims to identify host targets in different pathways where the phytocompounds of Kk possibly interact. **Materials and Methods:** Using the curated list of phytocompounds from 15 plants obtained from existing literature, each compound was searched against the Pubchem database and downloaded using which probable host targets were predicted in Swiss Target Prediction and Binding database. Probability scores of 0.5 and above was selected as high-confidence targets and a score of less than 0.5 as low-confidence targets. **Results:** We constructed a network of compounds and their possible interacting targets and identified three pathways, namely Phosphatidylinositol 3-Kinase (P13K/AKT), Repressor Activator Protein 1 (Rap1) and Mitogen Activated Protein Kinase (MAPK) pathways that the phytocompounds possibly modulate. Additionally, we performed molecular docking simulations for the phytocompounds with target's 3D structure to map the molecular interaction and ascertain their role in antiviral treatment, where we found ellagic acid as the best binder to the targets of these three pathways. **Conclusion:** Using a network pharmacology approach this study demonstrated that Kk can modulate three important molecular pathways (P13K/AKT, Rap1 and MAPK) associated with COVID-19 and this could be due to the effect of the phyto ingredient ellagic acid.

Keywords: COVID-19, Ellagic acid, *Kabasuk kudineer*, Molecular docking, Network Pharmacology.

Correspondence:

Dr. Ragothaman Madhava Rao Yennamalli

Department of Bioinformatics, School of Chemical and Biotechnology, SASTRA Deemed to be University, Thanjavur, Tamil Nadu, INDIA.

Email: ragothaman@scbt.sastra.edu

Dr. Sriram Sridharan

School of Chemical and Biotechnology, Centre for Advanced Research in Indian System of Medicine (CARISM), SASTRA (Deemed to be University), Thanjavur-613401, Tamil Nadu, INDIA.

Email: sriram@scbt.sastra.edu

Received: 18-02-2024;

Revised: 05-08-2024;

Accepted: 29-01-2025.

INTRODUCTION

A sudden outbreak of pneumonia, initially observed in the Wuhan city of China's Hubei province in December 2019, gradually became a novel strain of coronavirus-SARS-CoV-2, a positive stranded enveloped RNA virus belonging to the Corona viridae family. What started as a relatively unsuspecting epidemic slowly started spreading throughout the globe across countries and territories and was subsequently declared by the World Health Organization (WHO) as a pandemic on 11

March (<https://bestpractice.bmj.com/topics/en-gb/3000165>; <https://www.who.int/news-room/articles-detail/updated-who-advice-for-international-traffic-in-relation-to-the-outbreak-of-the-novel-coronavirus-2019-ncov-24-jan/>; <https://www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports>).

While, it is known that the Angiotensin-Converting Enzyme-2 (ACE2) is the preferred host receptor for the virus to bind and gain entry, the role of host proteins in the cytoplasm and in various organelles towards the virus life cycle, specifically in the replication, production and maturation are explained in detail here.¹ Recently, the SARS-CoV-2 and host protein interactome and the X-ray and cryo-EM complex of ACE2 with the Spike Glycoprotein (S Protein) were published, that showed the



DOI: 10.5530/ijper.20256024

Copyright Information :

Copyright Author (s) 2025 Distributed under Creative Commons CC-BY 4.0

Publishing Partner : Manuscript Technomedia. [www.mstechnomedia.com]

specificity of S protein for ACE2 was high with a binding affinity of 15 nM.^{2,3} This implies that targeting the S protein with small molecule inhibitors is relatively difficult because the inhibitor has to competitively bind to ACE2 and its binding affinity should be relatively higher than the spike protein.

SARS-CoV-2 and Its Pathogenesis

SARS-CoV-2, a member of the Betacoronavirus genus, exhibits a single-stranded positive-sense RNA genome of approximately 30 kilobases.⁴ This extensive genome is capable of encoding both structural and non-structural proteins critical to viral replication and infection. The virus utilizes its Spike (S) glycoproteins to attach to the Angiotensin-Converting Enzyme 2 (ACE2) receptors on host cells, a process further facilitated by the Host Transmembrane Protease Serine 2 (TMPRSS2).⁵ Upon entry, the viral RNA is translated into polyproteins, which are subsequently processed by viral proteases such as 3CLpro and PLpro to yield functional non-structural proteins that drive the replication and transcription machinery. This process generates subgenomic RNA for structural protein synthesis, leading to the assembly of virions that bud from the host cell.⁶⁻⁸

The pathogenesis of COVID-19 is closely tied to both the direct cytopathic effects of the virus and the host's immune response. Viral replication in the respiratory epithelium initiates a cascade of inflammatory signaling, leading to the recruitment of immune cells and the release of cytokines. Severe cases are often characterized by dysregulated immune responses, including cytokine storms that contribute to widespread tissue damage, Acute Respiratory Distress Syndrome (ARDS) and multi-organ failure.⁹ This dual aspect of viral and host-mediated pathology underscores the need for therapeutic strategies that target both the virus and the inflammatory processes it induces.

Therapeutic Targets Involved in SARS-CoV-2 Infection

One hallmark of severe COVID-19 cases is the cytokine storm, an exaggerated immune response marked by the excessive release of pro-inflammatory cytokines such as Interleukin-6 (IL-6), Tumor Necrosis Factor-Alpha (TNF- α) and interferons. This hyperactivation of the immune system can lead to systemic inflammation, endothelial dysfunction and organ failure.¹⁰⁻¹² Therapeutic approaches targeting the cytokine storm include IL-6 inhibitors like tocilizumab and JAK-STAT pathway inhibitors, which aim to restore immune homeostasis.

Potential therapeutic targets also include viral proteins essential for replication, such as RNA-dependent RNA Polymerase (RdRp), the main Protease (3CLpro) and Papain-like protease (PLpro).¹³ RdRp inhibitors like remdesivir mimic nucleotide substrates, halting viral RNA synthesis. Similarly, inhibitors targeting 3CLpro disrupt the processing of polyproteins necessary for viral assembly.¹⁴⁻¹⁶ These targets, alongside strategies to block the

ACE2-S protein interaction and prevent viral entry, represent a multifaceted approach to mitigating SARS-CoV-2 infection. By combining antiviral agents with therapies modulating the immune response, the severity and progression of COVID-19 can be effectively managed.

The list of interacting viral-host proteins, called as interactome, has been performed for many corona viridae family of viruses, such as SARS, MERS and most recently with COVID-19.¹⁷ From the viral entry inside the cell and up to its exit as mature virions, there are multiple host proteins that interact and aid in the production of viral particles. Specifically, it has been reported that apart from the structural proteins, the non-structural proteins have numerous interactions. An interactome data shows 27 discrete networks with intra-connectivity of some host proteins that are closely related.¹⁸ Thus, it is possible to identify pathways and host targets that get regulated after the virus enters the cell.

For managing and treating diseases like SARS-CoV-2, a multi-drug formulation that could bind with multiple targets associated with the disease and could exhibit a synergistic effect is the need of the hour. Here, the formulations from *Ayurveda* and *Siddha* become relevant as the formulations, in general, are multi-drug in nature working in a synergistic way. Hence, in the present study we have selected a natural *Siddha* formulation *Kabasurak kudineer* (Kk).

The clinical usage and dosage of Kk is prescribed in ancient Siddha texts. Specifically, Kk is used for treating *Kabasuram* (Swine flu), as mentioned in *Siddha* texts like *Theran karisal*, *Suravagadam*, *Yugi chinthamani* etc., with major symptoms that include body ache, fever, cough, fatigue, diarrhea, sore throat, shortness of breath and chest pain.¹⁹⁻²¹ The *Siddha* manuscript *Citta vaiittiyattirattu* (CVT) also mentions the usage of this formulation in treating *Aiyacuram* (Phlegmatic fevers). Previously Kk was effectively used to target host response during the outbreak of influenza virus (https://www.nhp.gov.in/swine-flu_mtl). 5 g of Kk powder can be mixed and boiled with 300 mL of water, reducing it to 30 mL and can be given with honey as vehicle, to the patients twice a day or as directed by the physician.²² This formulation is made up of 15 botanical ingredients (Table 1).

Network pharmacology or Poly-pharmacology is a technique that brings systems biology and computer-based virtual high-throughput screening together to study, understand, illustrate and visualize the action of multi-component drugs and formulations and the concerted interactions of their ingredients with multiple targets.^{66,67} This strategy could be used for better prediction of impending diseases and their subsequent preventive measures, to search and identify novel therapeutic targets, to re-purpose existing drugs and improve their safety and efficacy and to design new personalized drugs.^{68,69} Exploration of traditional poly-herbal formulations and their synergistic action against multiple targets associated with diseases is a

well-documented approach in Chinese medicine⁷⁰⁻⁸¹ and is recently gaining momentum with regard to Indian traditional system of medicine (*Ayurveda* and *Siddha*) and medicinal plants too.⁸²

With respect to Kk, recent reports talk about the *in silico* docking of phyto ingredients of Kk with the COVID-19 spike glycoprotein.⁸³ The studies report appreciable binding activity of the Kk active ingredients against the viral glycoprotein and both the studies have proposed that the effective action of the phytocompounds of Kk against the viral protein may be due to its immunomodulatory potential. However, there have been no reports on the molecular mechanistic action of Kk, its interaction, modulation or regulation of pathways and genes associated with SARS-CoV-2 till date to the best of our knowledge.

Hence, the present study is an attempt to carry out a network pharmacological approach to investigate the interactions between the phytoconstituents of Kk and multiple targets associated with SARS-CoV-2 and to comprehend the mechanistic action of the formulation.

MATERIALS AND METHODS

Materials

A network pharmacology model using the ingredients of *Kabasurak kudineer* and their bioactive compounds was developed using Cytoscape 3.2.1, a java based open-source software. The reported active compounds were created in 3D and saved in .sdf format. CORINA structure generator (<https://www.mn-am.com/products/corina>) was used to obtain the 3D structure of the phytocompounds. These were then queried in Binding DB (<https://www.bindingdb.org/>) database to get high ranked hits. These hits were then mapped to other target databases such as Uniprot, therapeutic target database and others.

Compound Screening

After the data collection from CVT and identifying the phytocompounds of the plants, they were searched against the PubChem database⁸⁴ to obtain the structure information in SMILES format. For those compounds having no structure

in PubChem, they were not used in constructing the network construction and were marked separately. The next task was to identify the potential targets in the human body which would interact with the phytocompounds. For this purpose, Swiss target prediction server (<https://www.swisstargetprediction.ch/>) was used.⁸⁵ The potential targets were extracted along with the probability score of the compound that might bind with a particular receptor. An arbitrarily selected value of 0.5 was chosen as the threshold and any target below this threshold were classified as low confidence targets. Targets with a score of 0.5 and above were classified as high confidence targets.

Network Construction

To visualize the interactions between phytocompounds and their respective target proteins, a network was constructed using Cytoscape, a widely used network visualization platform.⁸⁶ In this network, the phytocompounds and their targets were represented as nodes, while the interactions between them were depicted as edges. This approach facilitated a comprehensive view of the molecular relationships.

Following the network creation, the structures of the host target proteins were retrieved for further analysis. The UniProt database served as the primary resource for obtaining structural information. For targets with listed structures in UniProt, several filtering criteria were applied to ensure the selection of high-quality data. These criteria included:

Method of structure determination: Preference was given to structures determined using X-ray crystallography or cryo-electron microscopy due to their higher resolution.

Resolution: Only structures with resolution values within an acceptable range (typically <2.5 Å) were selected to ensure structural accuracy.

Sequence length: Targets with complete sequences matching the experimental data were prioritized to avoid partial or truncated structures.

Domain information: The presence of well-annotated domains relevant to the target's function was considered essential.

Table 1: Ingredients of *Kabasurak kudineer* and their pharmacological activity.

Sl. No.	Ingredient Name	Botanical name	Part(s) used	Quantity	No. of Phytocompounds	Pharmacological activity
1	Dry ginger/ <i>Chukku</i>	<i>Zingiber officinale</i> Roscoe	Rhizome	1	41 compounds. ²³	Dissolve phlegm associated with colds or chronic bronchitis, treat asthma and cough due to cold and coldness associated with shock. ²⁴
2	Long pepper/ <i>Thippili</i>	<i>Piper longum</i> L.	Fruit	1	Fruit and Root-23 compounds. ²⁵ Fruit-16 compounds. ²⁵	Decoction of immature fruits and roots is used in chronic bronchitis, cough and cold. ²⁵

Sl. No.	Ingredient Name	Botanical name	Part(s) used	Quantity	No. of Phytocompounds	Pharmacological activity
3	Cloves/Lavangam	<i>Syzygium aromaticum</i> (L.)	Flower bud	1	18 compounds ²⁶	Expectorant; Soothe certain respiratory conditions like cold, cough, asthma, bronchitis and sinusitis; helps in clearing the nasal tract. ²⁷
4	Climbing nettle roots/ <i>Sirukanjori</i>	<i>Tragia involucrata</i> L.	Root	1	8 compounds. ²⁸	Leaf-effective in treating pain and bronchitis; Root-treatment of high fever. ²⁹
5	Pellitory/ <i>Akkarakaram</i>	<i>Anacyclus pyrethrum</i> (L.) Lag.	Root	1	10 compounds. ³⁰	Akarkara root exhibits anticatarrhal properties, i.e. it expels old catarrh. ³⁰
6	<i>Barleria</i> roots/ <i>Neermulli ver</i>	<i>Hygrophila auriculata</i> (Schumach.) Heine	Root	1	2 compounds. ³¹	Tonic for asthma. ³¹
7	<i>Chebulic myrobalan/Kadukkai thole</i>	<i>Terminalia chebula</i> Retz.	Pericarp	1	13 compounds. ^{32,33}	Antitussive; ³⁴ Used in treatment of asthma, cough, dyspnea. ³⁵
8	<i>Adhatoda</i> root/ <i>Adhatoda ver</i>	<i>Justicia adhatoda</i> L.	Root	1	4 compounds. ³⁶⁻³⁸	Bronchodilatory activity; ³⁹ Used in treating chest and respiratory track infection; ⁴⁰ treatment of whooping cough, chronic bronchitis, asthma and excessive phlegm. ⁴¹
9	Indian borage/ <i>Karpooravalli</i>	<i>Coleus amboinicus</i> Lour.	Leaf	1	10 compounds. ⁴²⁻⁴⁵	Used in treatment of chronic coughs, asthma, bronchitis and sore throat; ⁴⁶ has bronchodilatory activity; ⁴⁷ Used to treat catarrhal infections; ⁴⁸ juice/decoction can be a worthy treatment for influenza, cough, bronchitis and throat problems. ⁴⁹
10	<i>Costus</i> root/ <i>Kottam</i>	<i>Saussurea costus</i> (Falc.) Lipsch.	Root	1	26 compounds. ⁵⁰	Roots are used mainly as an antispasmodic in asthma, cough. ⁵¹
11	<i>Tinospora</i> stem/ <i>Seenthil</i>	<i>Tinospora cordifolia</i> (Willd.) Hook.f. and Thomson.	Stem	1	39 compounds. ⁵²	Can be given as adjuvant therapy in chronic bronchitis patients in addition to standard treatment. ⁵³
12	Beetle killer root/ <i>Siruthekkku</i>	<i>Rothea serrata</i> (L.) Steane and Mabb.	Root	1	27 compounds. ⁵⁴	Root used in treatment of asthma. ⁵⁵
13	Nilavembu samoolam	<i>Andrographis paniculata</i> (Burm.f.) Nees	Whole plant	1	11 compounds. ⁵⁶	Good remedy as treatment for common respiratory infections such as colds and flu. ⁵⁷
14	Vattathiruppi Ver	<i>Cissampelos pareira</i> L.	Root	1	6 compounds. ⁵⁸⁻⁶³	Used in treatment of fever. ⁶⁴
15	Nut grass tubers/ <i>Koraikizhangu</i>	<i>Cyperus rotundus</i> L.	Rhizome	1	28 compounds. ⁶⁵	Tubers are used to treat cough, fever; Considered as an antispasmodic, antitussive; prescribed to treat bronchitis. ⁶⁵

This systematic process ensured the incorporation of reliable and high-quality structural data into the analysis, thereby enhancing the robustness of subsequent modeling and interpretation steps.

Molecular Modeling

For targets that did not have published structures, the Rosetta algorithm implemented in Robetta server⁸⁷ and Swissmodel server⁸⁸ was used for structure prediction, using a comparative modeling approach.

Assessing the quality of 3D modelled structures

The three-dimensional structures of the target proteins were generated using the Robetta and SwissModel servers.^{89,90} To ensure the reliability and accuracy of the predicted structures, a systematic quality assessment was conducted.

For models generated by the Robetta server, only those with a confidence score of 0.8 or higher were retained for further analysis. This threshold ensured the inclusion of high-quality models with robust structural predictions. Similarly, the SwissModel-generated structures were evaluated using their associated quality scores, including the Global Model Quality Estimation (GMQE) and QMEAN scores. Models with GMQE values above 0.7 and favorable QMEAN Z-scores, indicating

agreement with high-resolution experimental structures, were selected.

Additionally, structural integrity and stereochemical quality were validated using tools such as PROCHECK and MolProbity. These tools assessed parameters like Ramachandran plot distributions, bond angles and overall structural geometry, ensuring that the models adhered to standard protein structure quality benchmarks. This multi-step approach ensured that only the most reliable models were used for subsequent analysis.

Molecular Docking

To find out the interactions between the receptor and the ligand we did not specify the binding site but we used an unbiased approach of blind docking.^{91,92} In this we, defined the 3D grid map of the AutoGrid step such that 95% of the target is defined as the binding site. This unbiased approach has been shown to give better results. An exhaustive molecular docking approach-with 100 dockings for each phytochemical was performed using the lamarkian genetic algorithm in AutoDock 4.2. The compiled data obtained from molecular docking is listed in Supplementary Table S1. The overall lowest energy confirmation of the ligand, from all 100 docked confirmations was extracted and the coordinates were converted into a pdb file. This lowest energy confirmation-ligand was visualized along with the macromolecule in pymol and

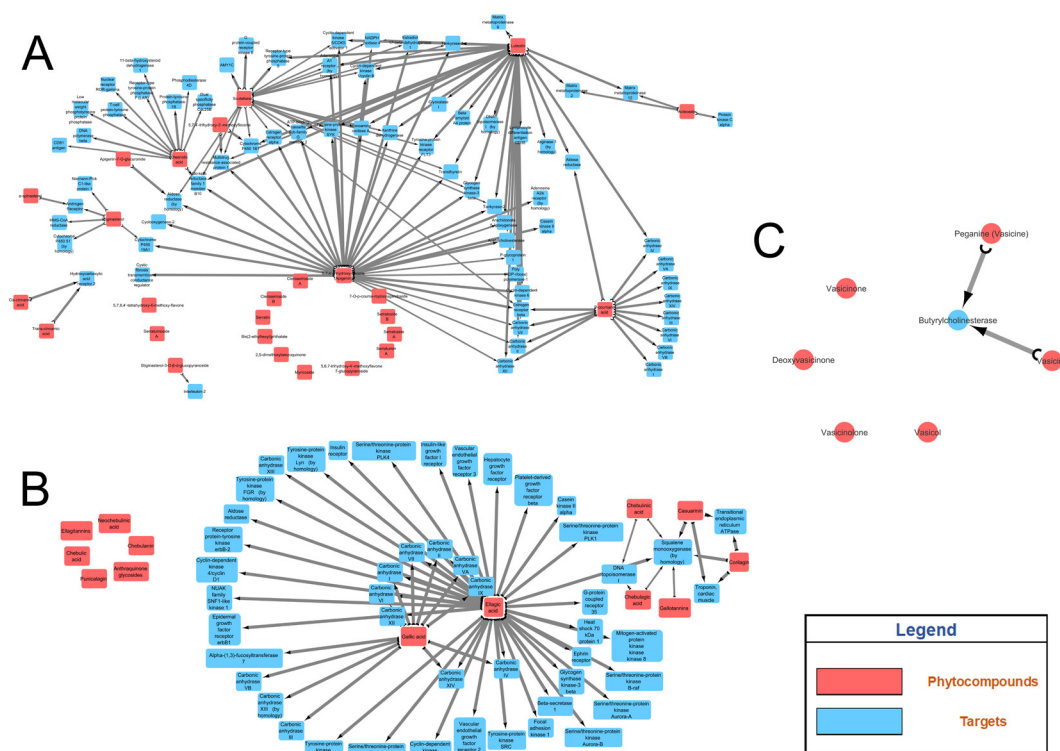


Figure 1: The networks of A) Beetle Killer root, B) *Chebulic myrobalan* and C) *Adhatoda* root the edges, represented in grey shows the interaction between a target and a phytochemical. The width of the edges depends on the potential binding probability score of the target with its phytochemical, the width increases as the probability score increases. The probability scores of potential targets for all the plants range from 0 to 1 classically, but after filtering them out, the high confidence targets range from 0.5 to 1. The plants have probability scores with their respective targets as following: A) Beetle Killer root-0.509 to 0.971, B) *Chebulic myrobalan*-0.874 to 1.000 and C) *Adatoda* root-1.000.

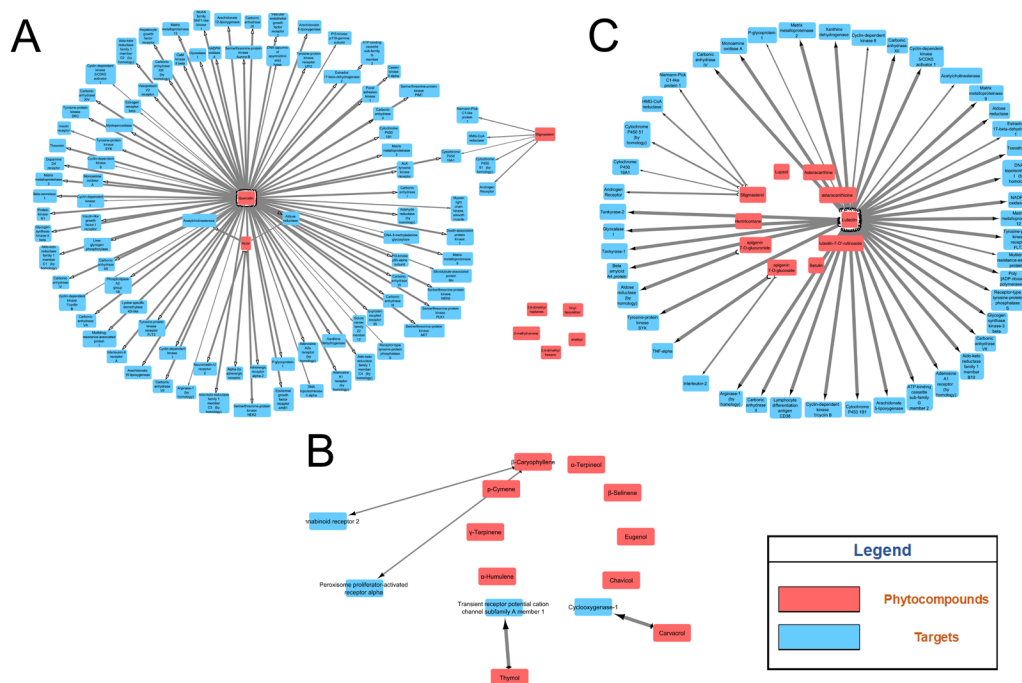


Figure 2: The networks of A) Climbing Nettle roots, B) Indian Borage and C) *Hygrophylla auriculata* (Schumach.) Heine. The edges, represented in grey shows the interaction between a target and a phytocompound. The width of the edges depends on the potential binding probability score of the target with its phytocompound, the width increases as the probability score increases. The probability scores of potential targets for all the plants range from 0 to 1 classically, but after filtering them out, the high confidence targets range from 0.5 to 1. The plants have probability scores with their respective targets as following: A) Climbing Nettle roots-0.514 to 1.000, B) Indian Borage-0.699 to 1.000 and C) *Hygrophylla auriculata* (Schumach.) Heine-0.514 to 1.000.

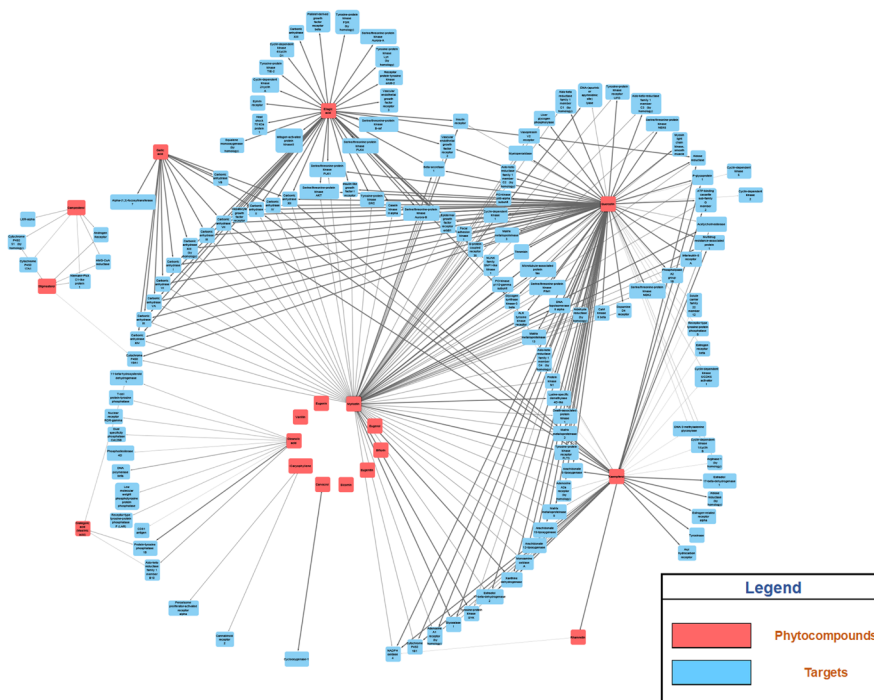


Figure 3: The network of Cloves. The edges, represented in grey shows the interaction between a target and a phytocompound. The width of the edges depends on the potential binding probability score of the target with its phytocompound, the width increases as the probability score increases. The probability scores of potential targets for all the plants range from 0 to 1 classically, but after filtering them out, the high confidence targets range from 0.5 to 1. The probability scores for this network ranged from 0.501 to 1.000.

the interactions mapped. We mapped pdb ids into uniprot ids by using tool, Retrieve/ID mapping (<https://www.uniprot.org/uploadlists/>).

KEGG Enrichment Analysis

We mapped pdb ids into uniprot ids by using tool, Retrieve/ID mapping (<https://www.uniprot.org/uploadlists/>). These target proteins were considered for pathway analysis in StringDB⁹³ and KEGG Mapper.⁹⁴ In StringDB, the confidence score for interactions were 0.4, 0.7 and 0.9. The proteins interactions as evidenced by gene fusion and experimental data were selected. The pathways of these proteins were analyzed further. The proteins were taken as input in KEGG Mapper, a tool to map pathways. In KEGG Mapper, we used the search pathway option and we selected “hsa”. We the proteins exported from StringDB were given as input, the important pathways were identified. The targets were selected for pathway reconstruction using Pajek software.⁹⁵

RESULTS

The current study involved a network pharmacological approach to understand the plausible molecular mechanistic action of Kk in the treatment of SARS-CoV-2 by studying the interactions between phytoconstituents of Kk and multiple targets associated with SARS-CoV-2. A network interaction was constructed

between the active ingredients of Kk, their multiple targets and the plausible pathways.

Network Pharmacology Scores

Adathoda root has about 6 phytoconstituents of which only two of them share the only possible high confidence target for this plant (Figure 1A), *Chebolic myrobalan* has 13 phytoconstituents and 48 targets (Figure 1B) and *Beetle killer* root has 27 phytoconstituents and 70 targets (Figure 1C). The plants have probability scores with their respective targets as following: *Beetle killer* root-0.509 to 0.971, *Chebolic myrobalan*-0.874 to 1.000 and *Adathoda* root-1.000.

Climbing nettle roots have about 8 phytoconstituents and 96 high confidence targets (Figure 2A), *Indian borage* has 10 phytoconstituents and 4 targets (Figure 2B) and *Hygrophilla auriculata* has 10 phytoconstituents and 44 targets (Figure 2C). The plants have probability scores with their respective targets as following: A) *Climbing nettle* roots-0.514 to 1.000, *Indian borage*-0.699 to 1.000 and *Hygrophilla auriculata*-0.514 to 1.000.

Clove (Figure 3) is the ingredient in this herbal concoction which has the greatest number of phytoconstituents with most interacting targets and hence the largest network with 18 phytoconstituents and 285 potential high confidence targets. The probability scores of potential targets for all the plants range from 0 to 1 classically,

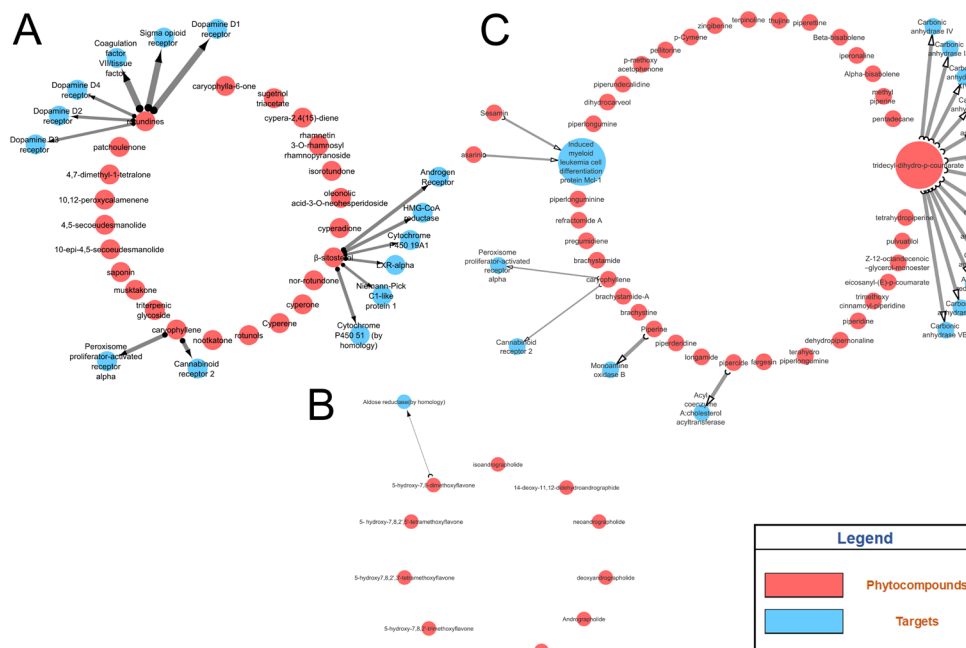


Figure 4: The networks of A) Nut Grass Tubers, B) *Nilavembu samoolam* and C) Long Pepper. The edges, represented in grey, show the interaction between a target and a phytoconstituent. The width of the edges depends on the potential binding probability score of the target with its phytoconstituent; the width increases as the probability score increases. The probability scores of potential targets for all the plants range from 0 to 1 classically, but after filtering them out, the high confidence targets range from 0.5 to 1. The plants have probability scores with their respective targets as following: A) Nut Grass Tubers-0.555 to 0.967, B) *Nilavembu samoolam*-0.522 and C) Long Pepper-0.699 to 1.000.

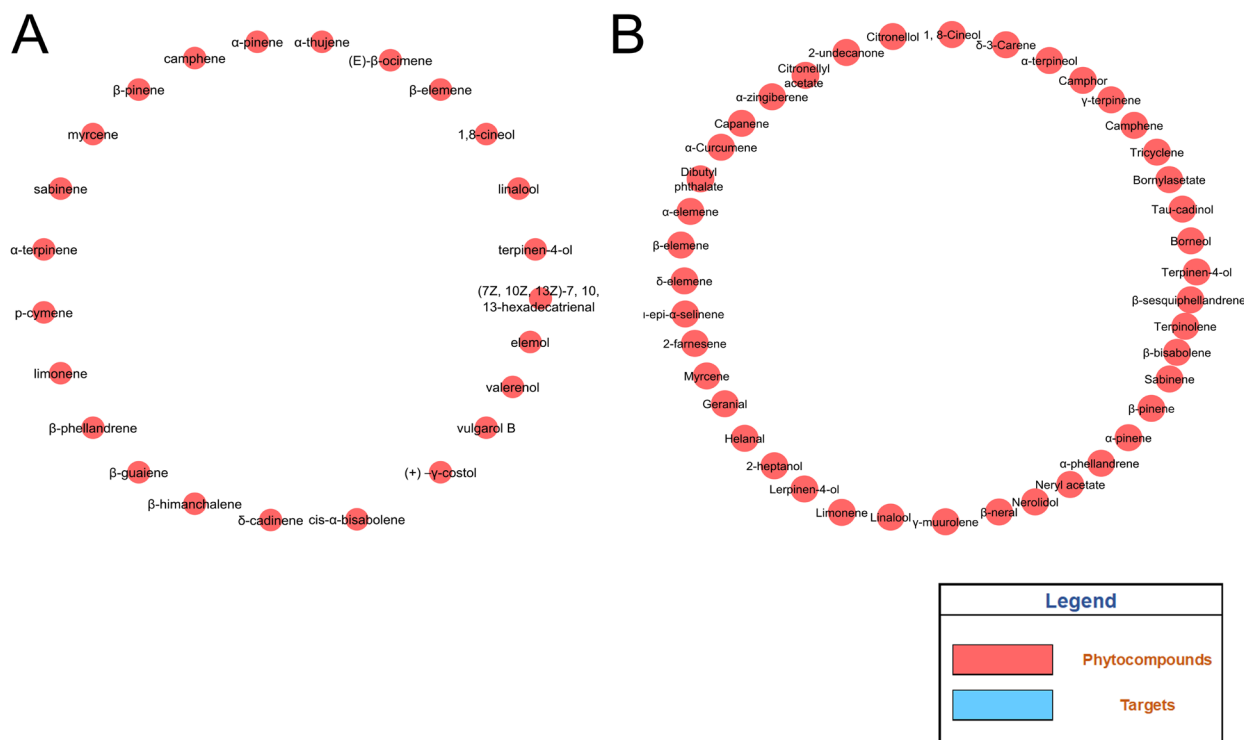


Figure 6: The networks of A) *Costus* root and B) Dry Ginger. The ingredients of the concoction put in this image have no potential targets so the networks without any edges representing only the phytochemicals. *Costus* root has 24 phytochemicals and the same is about 41 in the case of Dry Ginger.

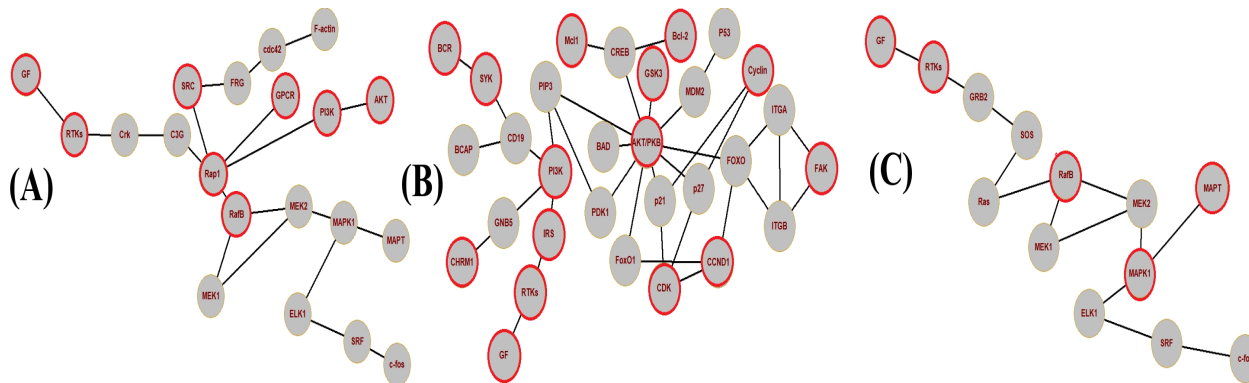


Figure 7: Pathways identified with KEGG Mapper. The proteins were mapped into three important pathways A) MAPK pathway B) PI3K/AKT pathway and Rap1 pathway. MAPK pathway, we have 18 proteins, colored in grey color. 8 proteins from our data have been highlighted. Similarly, PI3K-Akt pathway, we have 26 proteins, colored in grey color. 12 proteins from our data have been highlighted. PI3K-Akt pathway, we have 16 proteins, colored in grey color. 8 proteins from our data have been highlighted.

The coactive effects of *Siddha* medications with the conventional treatment have resulted in a promising improvement in the condition of COVID-19 patients.⁹⁷ Initial inflammatory stage of COVID-19 infection involved the administration of *Poorna chandirodayam* and *Gorojanai mathirai* along with other herbal and herbomineral *Siddha* formulations was performed to minimize hypoxia.⁹⁸ One research group have drawn the conclusion that *Nagaradi Kashaya* may be effective in battling SARS-CoV-2 and for performing additional clinical trials,

Nagaradi Kashaya may be a lead candidate.⁹⁹ To strengthen immunity against the COVID-19 infection, the AYUSH ministry suggested the herbal combination of *Ocimum tenuiflorum*, *Cinnamomum verum*, *Piper nigrum*, *Zingiber officinale* and *Vitis vinifera* as they were found to modulating the signalling of HIF-1, p53, PI3K-Akt, MAPK, cAMP, Ras, Wnt, NF-kappa B, IL-17, TNF and cGMP-PKG among other diverse pathways.¹⁰⁰ Recently, quite a few studies have been centered on *Kk*, a *Siddha* formulation made up of 15 botanical ingredients and a wide range

of pharmacological activities.²³⁻⁶⁵ Nine major active ingredients of the *Siddha* official formulation *Kk* were observed to be a potent inhibitor of SARS-CoV-2 spike protein and a new formulation called SNACK-V was proposed.⁸³ In one report, *Kk* used in the strategy against COVID-19 considerably decreased SARS-CoV-2 viral load among asymptomatic COVID-19 cases and did not record any negative effects but the pathways were not enriched and promising compound from *Kk* was not reported.¹⁰¹

In the present study, we constructed a network of compounds from *Kk* and their possible interacting targets and identified three pathways, namely P13K/AKT, Rap1 and MAPK pathways that the phytochemicals possibly modulate. Interestingly, all these 3 pathways are being looked on as possible therapeutic targets against COVID-19. All these 3 pathways when activated, result in the increased expression and release of pro-inflammatory mediators such as IL-6, TNF- α , IL-1 β and activation of factors like Activated Protein-1 (AP-1) and Nuclear Factor Kappa B (NF κ B)^{102,103} and there have been recent literature evidences that indicate an increased level of inflammatory mediators like cytokines and chemokines in COVID-19 patients.¹⁰⁴⁻¹⁰⁶

In addition, activated P13K/AKT and MAPK pathways have been found to be associated with viral endocytosis¹⁰⁷ and lung tissue fibrosis^{108,109} especially in COVID-19 patients. A clathrin-mediated pathway regulated by activated P13K/AKT seems to facilitate the endocytosis of SARS-CoV-2, leading to reduction of ACE2 and a subsequent increase in the levels of inflammatory cytokines finally causing lung fibrosis. Similarly, it has been shown that vital thrombotic events such as platelet aggregation, arterial thrombosis and endothelial dysfunction observed in COVID-19 patients may be due to disproportionately activated p38 MAPK pathway that may in turn lead to apoptosis, impaired contraction and fibrosis.¹¹⁰

The Rap1 signaling pathway has been identified as one of the prominent pathways involved in the regulation of immune system. The Rap1 pathway activates 3 different secondary messengers namely 3',5' Cyclic Adenosine Monophosphate (cAMP), Calcium Ion (Ca²⁺) and Diacylglycerol (DAG) that are required for cell position signaling in viral infections.^{82,111} The activation of cAMP triggers a cascade reaction activating cAMP dependent Protein Kinase-A (PKA) and an Exchange Protein Activated by cAMP (EPAC). It is also involved in the opening of ion channels that help in regulation of Ca²⁺ levels and thereby controls T cell proliferation and cytokine production.^{112,113} The cAMP/PKA pathway has been found to modulate the proliferation and transcription of cytokine genes like TNF- α , IFN- γ and Interleukins.¹¹⁴ cAMP is found to have dual and contrasting roles with regard to viral infection where on one hand it could decrease viral entry and replication and on the other hand it could suppress antiviral immune responses. Hence therapeutic medications that activate cAMP could act as immunosuppressants and those which repress cAMP signaling could act as immunostimulators.¹¹³

Based on the results obtained in this study and the available information on the 3 pathways, it could be safely hypothesized that the plausible molecular mechanistic action of *Kk* in the management and treatment of SARS-CoV-2 could be through inhibition of P13K/AKT, MAPK and Rap1 signaling pathways, that would in turn result in reduction of expression and release of pro-inflammatory cytokines, viral endocytosis and lung fibrosis all of which are associated with COVID-19. In the present study, among all the phytoconstituents that interacted with multiple protein targets, Ellagic acid was found to have the best binding affinity [Supplementary Table S1]. Ellagic acid is a polyphenol that is found mainly in fruits. The ability of Polyphenols' potential to treat cancer by blocking the PI3K/Akt/mTOR signaling pathway has been reported,¹¹⁵ and the compound ellagic acid has been reported to show anti-inflammatory and anti-cancer activity.¹¹⁶⁻¹¹⁸ Interestingly, Ellagic acid is currently being put forth as a possible therapeutic agent against COVID-19 based on its proven therapeutic ability. The antiviral effect of Ellagic acid against Influenza A (H3N2), Rhinoviruses (HRV-2,3 and 4), Ebola, HIV-1, HSV-1 and noroviruses have been well documented.¹¹⁹⁻¹²¹

Ellagic acid's protective effect against lung damage could be attributed to its excellent anti-oxidant potential and it was also found to possess anti-inflammatory activity and could bring down the levels of IL-6.¹²² In addition, Ellagic acid was also found to exhibit synergistic action with antimalarial drugs that are currently being prescribed as a treatment strategy against COVID-19.¹²³ In another recent study carried out by Usama Ramadan *et al.* 2020,¹²⁴ when more than 496 phenolic compounds were virtually screened against the active site of SARS-CoV Main protease, it was found that only Ellagic acid along with acetylglucopetunidin and isoxanthohumol obeyed the Lipinski's and Veber's rules of drug-likeness and possessed higher degree of bioavailability.

CONCLUSION

There have been multiple reports about the *Siddha* formulation *Kk* being effective in preventing and curing the viral disease. However, there is a paucity of studies performed to identify the molecular level mechanism of *Kk* and its effectiveness with respect to COVID-19. We hope that this study fills that gap and also shows that using computational techniques such as network pharmacology and molecular docking, potential phytochemicals and their most probable host targets can be identified. Specifically, we identified the role of *Kk* phytochemicals and their interaction with 3 major pathways namely the Phosphatidylinositol 3-Kinase (P13K/AKT), Mitogen Activated Protein Kinase (MAPK) and Repressor Activator Protein 1 (Rap1) signaling pathways. It has to be noted that these three pathways are potential therapeutic targets against COVID-19 as reported in literature. Also, that the high specific binding of Ellagic acid, using a blind molecular docking approach, is statistically significant. This study clearly

illustrates that, traditional formulations like Kk with specific active ingredients could prove to be beneficial as an alternative source of therapeutic intervention both in the treatment and prevention of COVID-19. Further wet lab experiments and clinical trials could confirm and establish the effectiveness of this formulation against COVID-19.

ACKNOWLEDGEMENT

We acknowledge SASTRA (Deemed to be University) for the Infrastructure support provided to carry out this research work.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ABBREVIATIONS

Kk: *Kabasura kudineer*; **SMILES:** Simplified Molecular Input Line Entry System; **DB:** Database; **P13K/AKT:** Phosphatidylinositol 3-kinase; **MAPK:** Mitogen activated protein kinase; **Rap1:** Repressor activator protein 1; **ACE2:** Angiotensin-converting enzyme-2; **SARS:** Severe acute respiratory syndrome; **MERS:** Middle East respiratory syndrome; **KEGG:** Kyoto Encyclopedia of Genes and Genomes; **EPAC:** Exchange protein activated by cAMP.

CREDIT AUTHOR STATEMENT

Brindha Pemaiah, Ragothaman M. Yennamalli, Sriram Sridharan and Davidraj Chellappan conceived the idea and designed the experiments; Yatindrapravanan Narasimhan and Shashank Ravichandran performed the molecular docking and draft of the figures, Krishna Kant Gupta performed the network analysis and wrote the first manuscript draft; All authors were involved in editing and finalizing the manuscript.

SUMMARY

Kabasura kudineer (Kk) is a classical *Siddha* medicine used for the management of many types of fevers and flu with respiratory complications. In this study, a network pharmacological approach has been employed to investigate the interactions between the phytoconstituents of Kk and multiple targets associated with SARS-CoV-2 and also obtain valuable inputs on the mechanism of action of the formulation. The present study indicates that Kk could modulate three important molecular Pathways (P13K/AKT, Rap1 and MAPK) associated with COVID-19 and this could be due to the effect of one of its phytoingredient, Ellagic acid.

REFERENCES

- Ni W, Yang X, Yang D, et al. Role of angiotensin-converting enzyme 2 (ACE2) in COVID-19. *J. Crit. Care.* 2020;24:422. doi:10.1186/s13054-020-03120-0
- Gordon DE, Jang GM, Bouhaddou M, et al. A SARS-CoV-2 protein interaction map reveals targets for drug repurposing. *Nature.* 2020;583:459-68. doi:10.1038/s41586-020-2286-9
- Lan J, Ge J, Yu J, et al. Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. *Nature.* 2020;581:21520. doi:10.1038/s41586-020-2180-5
- Kirchdoerfer RN, Cottrell CA, Wang N, Pallesen J, Yassine HM, Turner HL, Corbett KS, Graham BS, McLellan JS, Ward AB. *Nature.* 2016;531:118
- Xia M, Liu C, Wang W, Xu Q, Lan S, Feng F, et al. *Cell Res.* 2020;30:343
- Muthumani K, Falzarano D, Reuschel EL, Tingey C, Flingai S, Villarreal DO, et al. *Sci. Transl. Med.* 2015;7:301ra132.
- Jiang S, He Y, Liu S. *Emerg. Infect. Dis.* 2005;11:1016
- Yang Z, Kong W, Huang Y, Roberts A, Murphy BR, Subbarao K, Nabel GJ. *Nature.* 2004;428:561
- Chen CM, Lu HC, Tung YT, Chen W. *Biomedicines.* 2020;8:230.
- Zhang H, Baker A. *Crit Care.* 2017;21:305
- Ziebuhr J. *Curr. Opin. Microbiol.* 2004;7:412.
- Prentice E, McAuliffe J, Lu X, Subbarao K, Denison MR. *J.Virol.* 2004;78:9977.
- Ansari MA, Jamal QMS, Rehman S, Almatroudi A, Alzohairy MA, Alomary MN, Tripathi T, Alharbi AH, Adil SF, Khan M. *Arab. J. Chem.* 2020;13:8069.
- Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, Ruan L, Song B, Cai Y, Wei. *N. Engl. J. Med.* 2020;382:1787.
- Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, Shi Z, Hu Z, Zhong W, Xiao G. *Cell Res.* 2020;30:269.
- Li G, De Clercq E. *Nat Rev Drug Discov* 2020;19:149.
- Lim Y, Ng Y, Tam J, Liu D. Human Coronaviruses: A Review of Virus-Host Interactions. *J. Dis.* 2016;4(3):26. doi:10.3390/diseases4030026
- Brito AF, Pinney JW. Protein-Protein Interactions in Virus-Host Systems. *Front. Microbiol.* 2017;8. doi:10.3389/fmicb.2017.01557
- Kuppusamy Mudaliar, KN. Suram, *Siddha Maruthuvam*, Tamil Nadu *Siddha Maruthuvavariyam.*1987
- Murugesha Mudaliar, KS. *Siddha Materia Medica*, Directorate of Indian Medicine and Homeopathy, Chennai; 2013
- Uthamarayan, KS. *Siddha Maruthuvangachurukkam*, Directorate of Indian Medicine and Homeopathy, Chennai; 2006
- Siddha Formulary of India*, Part-I, 1st ed. Ministry of Health and Family Welfare, Govt of India, New Delhi; 1992.
- Fei-fei Qin, Hui-lian Xu. Active compounds in Gingers and their therapeutic use in Complimentary medication. *MAPSB.* 2008;2(2):72-8.
- Bensky, D., Gamble, A., 1986. Chinese herbal medicine, *Materia medica Eastland Press*, Seattle, WA, pp. 562.
- Maitreyi Zaveri et al. Chemistry and Pharmacology of *Piper longum* L. *Int. J. Pharm. Sci. Rev. Res.* 2010;5(1):67-76.
- Cortés-Rojas DF, de Souza CRF, Oliveira WP. Clove (*Syzygium aromaticum*): a precious spice. *Asian Pac. J. Trop. Biomed.* 2014;4(2):90-6. doi:10.1016/s2221-1691(14)60215-x
- Parle Milind, Khanna Deepa. CLOVE: A Champion Spice. *Int. J. Ayurveda Res.* 2011;2(1):47-54.
- Panda D, Santhosh Kumar, D, Gouri Kumar D. Phytochemical Examination and Antimicrobial activity of various solvent extracts and the selected isolated compounds from roots of *Tragia involucrata* Linn. *Int. J. Pharm. Sci. Drug Res.* 2012;4:44-8.
- Kirtikar KR, Basu BD. 1987. Indian medicinal plants. Second ed. Delhi, pp. 757-9.
- Bhuktar AS.2000. Plant Resources Development, In: Mungikar Am, Bhuktar AS, editor. Sarswati Printing Press; Aurangabad, pp. 146-52
- Usmani A, Khushtar M, Arif M, Siddiqui Mohd, Sing S, Mujahid M. Pharmacognostic and phytopharmacology study of *Anacyclus pyrethrum*: An insight. *J. Appl. Pharm. Sci.* 2016;144-50. doi:10.7324/japs.2016.60325
- Afshari AR, Sadeghnia HR, Mollazadeh H. A Review on Potential Mechanisms of *Terminalia chebula* in Alzheimer's Disease. *Adv Pharmacol Sci.* 2016;1-14. doi:10.1155/2016/8964849
- Juang LJ, Sheu SJ, Lin TC. Determination of hydrolyzable tannins in the fruit of *Terminalia chebula* Retz. by high-performance liquid chromatography and capillary electrophoresis. *J. Sep. Sci.* 2004;27(9):718-24. doi:10.1002/jssc.200401741
- Kokate CK, Purohit, AP, Gokhale SP. Pharmacognosy. 2001 12th Edn., Nirali Prakashan, Pune; pp. 216-2'17.
- Nadkarni KM Indian Material Medica. Popular Prakashan Pvt. Ltd., 1997 Bombay; pp. 1202-1211.
- Amin AH, Mehta DR. A Bronchodilator Alkaloid (Vasicinone) from *Adhatoda vasica* Nees. *Nature.* 1959;184(4695):1317. doi:10.1038/1841317a0
- Gupta OP, Sharma ML, Ghatak BJ, Atal CK. Pharmacological investigations of vasicinone and vasicinone--the alkaloids of *Adhatoda vasica*. *Indian J Med Res.*1977;66(4): 680-91.
- Ajay Sharma., Garima Bhardwaj., Damanjit Singh Cannoo. Overview of Phytochemistry and Pharmacology of *Adhatoda vasica*. *Int. J. Adv. Manag. Technol. Eng. Sci* 2018;8(3).
- Al-Shamma A, Drake S, Flynn DL, et al. Antimicrobial Agents from higher plants. Antimicrobial agents from *Peganum harmala* seeds. *J. Nat. Prod.* 1981;44(6):745-7. doi:10.1021/np50018a025

40. Maikhuri RK, Gangwar AK. Ethnobiological notes on the Khasi and Garo tribes of Meghalaya, Northeast India. *Econ. Bot.* 1993;47(4):345-57. doi:10.1007/bf02907348
41. Pushpangadan P, Nyman U, George V. Glimpses of Indian Ethnopharmacology. 1995 Tropical Botanic Garden and Research Institute, Kerala; 309-83.
42. Menéndez Castillo RA, Pavón González V. *Plectranthus amboinicus* (Lour.) Spreng. *Rev. Cuba. Plantas Med.* 1999;4(3):110-5.
43. Singh G, Singh OP, Prasad YR, de Lampasona MP, Catalan C. Studies on essential oils, Part 33: chemical and insecticidal investigations on leaf oil of *Coleus amboinicus* Lour. *Flavour Fragr. J.* 2002;17(6):440-2. doi:10.1002/ffj.1123
44. Senthilkumar A, Venkatesalu V. Chemical composition and larvicidal activity of the essential oil of *Plectranthus amboinicus* (Lour.) Spreng against *Anopheles stephensi*: a malarial vector mosquito. *Parasitol. Res.* 2010;107(5):1275-78. doi:10.1007/s00436-010-1996-6
45. Dutta S. Essential oil of *Coleus aromaticus* of Indian origin. *Indian Oil and Soap J.* 1959;25:120.
46. Jain SK, Lata S. Amazonian uses of some plants growing in India. *Indig. Knowl. Dev. Monit* 1996;4, 21-23.
47. Carbajal D et al. Pharmacological screening of plant decoctions commonly used in Cuban folk medicine. *J. Ethnopharmacol.* 1991;33, 21-24. https://doi.org/10.1016/0378-8741(91)90155-7
48. Cano JH, Volpato G. Herbal mixtures in the traditional medicine of Eastern Cuba. *J. Ethnopharmacol.* 2004;90, 293-316. https://doi.org/10.1016/j.jep.2003.10.012
49. Cartaxoa SL, Souzaa MMA, Albuquerque UP. Medicinal plants with bioprospecting potential used in semi-arid northeastern Brazil. *J. Ethnopharmacol.* 2010;131:326-42. https://doi.org/10.1016/j.jep.2010.07.003
50. Andola H, Lohani H, Chauhan N, Gwari G, Bhandari U. Volatile constituents of *Saussurea costus* roots cultivated in Uttarakhand Himalayas, India. *Pharmacognosy Res.* 2013;5(3):179. doi:10.4103/0974-8490.112424
51. Chopra RN, Nayar SL, Chopra IC. Glossary of Indian Medicinal Plants. Publication and Information Directorate, CSIR; 1956.
52. Sharma P, Dwivedee BP, Bisht D, Dash AK, Kumar D. The chemical constituents and diverse pharmacological importance of *Tinospora cordifolia*. *Heliyon.* 2019;5(9):e02437. doi:10.1016/j.heliyon.2019.e02437
53. Ratan Kumar. A randomized study of effect of *Tinospora Cordifolia* in chronic bronchitis patients. *IOSR J. Med. Dent. Sci. (IOSR-JDMS).* 2018;17(7):5-10. DOI: 10.9790/0853-1707160510
54. Wang JH, Luan F, He XD, Wang Y, Li MX. Traditional uses and pharmacological properties of *Clerodendrum* phytochemicals. *J. Tradit. Complement. Med.* 2018;8(1):24-38. doi:10.1016/j.jtcm.2017.04.001
55. Singh Mukesh K. *Clerodendrum serratum*: A clinical approach. *J. Appl. Pharm. Sci.* 2012;2(2):11-15.
56. Chao WW, Lin BF. Isolation and identification of bioactive compounds in *Andrographis paniculata* (Chuanxinlian). *Chin. Med.* 2010;5(1):17. doi:10.1186/1749-8546-5-17
57. Thamlikitkul V, Dechatiwongse T, Theerapong S, et al. Efficacy of *Andrographis paniculata*, Nees for pharyngotonsillitis in adults. *J Med Assoc Thai.* 1991;74(10):437-42.
58. Barbosa-Filho J, Da-Cunha EVL, Cornélio ML, Silva Dias CD, Gray AI. Cissaglaberrimine, an aporphine alkaloid from *Cissampelos glaberrima*. *Phytochem.* 1997;44(5):959-61. doi:10.1016/s0031-9422(96)00689-9
59. Lee SS, Lin YJ, Chen CK, Liu KCS, Chen CH. Quaternary Alkaloids from *Litsea cubeba* and *Cryptocarya konishii*. *J. Nat. Prod.* 1993;56(11):1971-6. doi:10.1021/np50101a016
60. Anwer F, Popli SP, Srivastava RM, Khare MP. Studies in medicinal plants. Part III. Protoberberine alkaloids from the roots of *Cissampelos pareira* linn. *Experientia.* 1968;24(10):999. doi:10.1007/bf02138704
61. Mambu L, Martin MT, Razafimahefa D, Ramanitrahambola D, Rasoanaivo P, Frappier F. Spectral characterisation and Antiplasmodial activity of Bisbenzyl isoquinolines from *Isolona ghesquiereina*. *Planta Med.* 2000;66(6):537-40. doi:10.1055/s-2000-8610
62. Bhatnagar AK, Bhattacharji S, Roy AC, Popli SP, Dhar ML. Chemical examination of the roots of *Cissampelos pareira*. IV. Structure and stereochemistry of hayatin. *The J. Org. Chem.* 1967;32(3):819-20. doi:10.1021/jo01278a071
63. Azucena GC et al. Antileishmanial, antitrypanosomal and cytotoxic screening of ethnopharmacologically selected Peruvian plants. *Parasitol Res.* 2012;110:1381-92. https://doi.org/10.1007/s00436-011-2638-3
64. Reza HM, Shohel M, Aziz SB, et al. Phytochemical and Pharmacological Investigation of Ethanol Extract of *Cissampelos pareira*. *Indian J. Pharm. Sci.* 2014;76(5):455-8. Accessed May 25, 2022.
65. Sultana S, Ali M, Mir SR. Chemical Constituents from the Rhizomes of *Cyperus Rotundus* L. *The Open Plant Sci.* 2017;10(1):82-91. doi:10.2174/1874294701710010082
66. Hopkins AL. Network pharmacology. *Nat. Biotechnol.* 2007;25(10):1110-1. doi:10.1038/nbt1007-1110
67. Patwardhan B. Editorial: The New Pharmacognosy. *Combinatorial Chemistry and High Throughput Screening* 2014;17(2):97. doi:10.2174/138620731702140119160627
68. Hopkins AL. Network pharmacology: the next paradigm in drug discovery. *Nat. Chem. Biol.* 2008;4(11):682-90. doi:10.1038/nchembio.118
69. Cheng F, Liu C, Jiang J, et al. Prediction of Drug-Target Interactions and Drug Repositioning via Network-Based Inference. *Altman RB, ed. PLoS Comput. Biol.* 2012;8(5):e1002503. doi:10.1371/journal.pcbi.1002503
70. Zhang G, Li Q, Chen Q, Su S. Network Pharmacology: A New Approach for Chinese Herbal Medicine Research. *eCAM.* 2013;2013:1-9. doi:10.1155/2013/621423
71. Li S, Zhang B. Traditional Chinese medicine network pharmacology: theory, methodology and application. *Chin. J. Nat. Med.* 2013;11(2):110-20. doi:10.1016/s1875-5364(13)60037-0
72. Tao W, Xu X, Wang X, et al. Network pharmacology-based prediction of the active ingredients and potential targets of Chinese herbal *Radix Curcumae* formula for application to cardiovascular disease. *J. Ethnopharmacol.* 2013;145(1):1-10. doi:10.1016/j.jep.2012.09.051
73. Wang Y, Liu Z, Li C, et al. Drug Target Prediction Based on the Herbs Components: The Study on the Multitargets Pharmacological Mechanism of Qishenkeli Acting on the Coronary Heart Disease. *eCAM.* 2012;2012:1-10. doi:10.1155/2012/698531
74. Zhang XL, Xing RG, Chen L, Liu CR, Miao ZG. PI3K/Akt signaling is involved in the pathogenesis of bleomycin-induced pulmonary fibrosis via regulation of epithelial-mesenchymal transition. *Mol. Med. Rep.* 2016;14(6):5699-5706. doi:10.3892/mmr.2016.5960
75. Tang J, Aittokallio T. Network Pharmacology Strategies Toward Multi-Target Anticancer Therapies: From Computational Models to Experimental Design Principles. *Curr. Pharm. Des.* 2014;20(1):23-36. doi:10.2174/13816128113199990470
76. Li J, Lu C, Jiang M, et al. Traditional Chinese Medicine-Based Network Pharmacology Could Lead to New Multicompound Drug Discovery. *eCAM;* 2012;2012:1-11. doi:10.1155/2012/149762
77. Li S. Framework and practice of network-based studies for Chinese herbal formula. *Chin. J. Integr. Med.* 2007;5(5):489-93. doi:10.3736/jcim20070501
78. Gu J, Gui Y, Chen L, Yuan G, Lu HZ, Xu X. Use of Natural Products as Chemical Library for Drug Discovery and Network Pharmacology. *D Cox, ed. PLoS One.* 2013;8(4):e62839. doi:10.1371/journal.pone.0062839
79. Pei L, Bao Y, Liu S, Zheng J, Chen X. Material Basis of Chinese Herbal Formulas Explored by Combining Pharmacokinetics with Network Pharmacology. *S Karnik, ed. PLoS One.* 2013;8(2):e57414. doi:10.1371/journal.pone.0057414
80. Zhao Y, Wang M, Tsering J, et al. An Integrated Study on the Antitumor Effect and Mechanism of Triphala Against Gynecological Cancers Based on Network Pharmacological Prediction and *in vitro* Experimental Validation. *Integr. Cancer Ther.* 2018;17:894-901. https://doi.org/10.1177/1534735418774410
81. Xu H, Zhang Y, Lei Y, et al. A Systems Biology-Based Approach to Uncovering the Molecular Mechanisms Underlying the Effects of Dragon's Blood Tablet in Colitis, Involving the Integration of Chemical Analysis, ADME Prediction and Network Pharmacology. *Baak JPA, ed. PLoS One.* 2014;9(7):e101432. doi:10.1371/journal.pone.0101432
82. Khanal P, Dey YN, Patil R, et al. Combination of system biology to probe the anti-viral activity of andrographolide and its derivative against COVID-19. *RSC Adv.* 2021;11(9):5065-79. doi:10.1039/d0ra10529e
83. Kiran G, Karthik L, Shree Devi MS, et al. *In Silico* computational screening of *Kabasura Kudineer* - Official *Siddha* Formulation and JACOM against SARS-CoV-2 spike protein. *J. Ayurveda Integr. Med.* 2022;13(1):100324. doi:10.1016/j.jaim.2020.05.009
84. Kim S, Chen J, Cheng T, et al. PubChem in 2021: New data content and improved web interfaces. *Nucleic Acids Res.* 2021;49(D1):D1388-D1395. doi:10.1093/nar/gkaa971
85. Gfeller D, Grosdidier A, Wirth M, Daina A, Michielin O, Zoete V. Swiss Target Prediction: a web server for target prediction of bioactive small molecules. *Nucleic Acids Res.* 2014;42(W1):W32-W38. doi:10.1093/nar/gku293
86. Su G, Morris JH, Demchak B, Bader GD. Biological network exploration with cytoscape 3. *Current protocols in bioinformatics / editorial board andreas D Baxevas [et al].* 2014;47:8.13.1-8.13.24. doi:10.1002/0471250953.bi0813s47
87. Kim DE, Chivian D, Baker D. Protein structure prediction and analysis using the Robetta server. *Nucleic Acids Res.* 2004;32 (Web Server): W526-W531. doi:10.1093/nar/gkh468
88. Waterhouse A, Bertoni M, Bienert S, et al. Swiss-Model: Homology modelling of protein structures and complexes. *Nucleic Acids Res.* 2018;46(W1):W296-W303. doi:10.1093/nar/gky427
89. Zrieq R, Snoussi M, Algahtan FD, Tasleem M, Saeed M, Noumi E, et al. Repurposing of anisomycin and oleandomycin as a potential anti-(SARS-CoV-2) virus targeting key enzymes using virtual computational approaches. *Cellular and Molecular Biology.* 2022;67(5):387-398. https://doi.org/10.14715/cmb/2021.67.5.51
90. Zrieq R, Ahmad I, Snoussi M, Noumi E, Iriti M, Algahtani FD, Patel H, Saeed M, Tasleem M, Sulaiman S, et al. Tomatidine and Patchouli Alcohol as Inhibitors of SARS-CoV-2 Enzymes (3CLpro, PLpro and NSP15) by Molecular Docking and Molecular Dynamics Simulations. *International Journal of Molecular Sciences.* 2021;22(19):10693. https://doi.org/10.3390/ijms221910693
91. Csaba Hetényi, David van der Spoel, Blind docking of drug-sized compounds to proteins with up to a thousand residues. *FEBS Letters.* 2006;580 (5):1447-50.
92. Hetényi C, Van der Spoel D. Efficient docking of peptides to proteins without prior knowledge of the binding site. *Protein Science.* 2002;11:1729-37. https://doi.org/10.1110/ps.0202302
93. Szklarczyk D, Gable AL, Lyon D, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide

- experimental datasets. *Nucleic Acids Res.* 2019; 47(Database issue): D607-13. doi:10.1093/nar/gky1131
94. Aoki-Kinoshita KF, Kanehisa M. KEGG Primer: An Introduction to Pathway Analysis Using KEGG. *NCI_PID.* 2007; doi:10.1038/pid.2007.2
 95. Pavlopoulos GA, Wegener AL, Schneider R. A survey of visualization tools for biological network analysis. *BioData Min.* 2008;1(1). doi:10.1186/1756-0381-1-12
 96. Jain J, Kumar A, Narayanan V, Ramaswamy RS, Sathiyarajeswaran P, Devi MS, et al. Antiviral activity of ethanolic extract of *Nilavembu Kudineer* against dengue and chikungunya virus through *in vitro* evaluation. *J Ayurveda Integr Med.* 2019;18:30073-1, 10.1016/j.jaim.2018.05.006
 97. Chitra SM, Mallika P, Anbu N, Narayanaswamy R, Sugunabai A, David Paul Raj RS, et al. An open clinical evaluation of selected Siddha regimen in expediting the management of COVID-19 - a randomized controlled study. *J Ayurveda Integr Med.* 2022;13(1):100397. doi: 10.1016/j.jaim.2021.01.002.
 98. Jeyavenkatesh J, Saravanapandian P, Amali JancyMargaret M, Shanmuga Priya R. Effect of *Siddha* medicine *Poorna chandirodayam* and *Gorojanai mathirai* among Covid 19 patients suffering from hypoxia - A case series. *J Ayurveda Integr Med.* 2022;13(2):100553. doi: 10.1016/j.jaim.2022.100553.
 99. Gandhi AJ, Rupareliya JD, Shukla VJ, Donga SB, Acharya R. An *Ayurvedic* perspective along with *in silico* study of the drugs for the management of SARS-CoV-2. *J Ayurveda Integr Med.* 2022;13(1):100343. doi: 10.1016/j.jaim.2020.07.002.
 100. Khanal P, Duyu T, Patil BM, Dey YN, Pasha I, Wanjari M, et al. Network pharmacology of AYUSH recommended immune-boosting medicinal plants against COVID-19. *J Ayurveda Integr Med.* 2022;13(1):100374. doi: 10.1016/j.jaim.2020.11.004.
 101. Natarajan S, Anbarasi C, Sathiyarajeswaran P, Manickam P, Geetha S, Kathiravan R et al. *Kabasura Kudineer* (KSK), a poly-herbal Siddha medicine, reduced SARS-CoV-2 viral load in asymptomatic COVID-19 individuals as compared to vitamin C and zinc supplementation: findings from a prospective, exploratory, open-labeled, comparative, randomized controlled trial, Tamil Nadu, India. *Trials.* 2021;22(1):623. doi: 10.1186/s13063-021-05583-0.
 102. Yodkeeree S, Ooppachai C, Pompimon W, Limtrakul (Dejkriengkraikul) P. O-Methylbulbocapnine and Dicentrine Suppress LPS-Induced Inflammatory Response by Blocking NF- κ B and AP-1 Activation through Inhibiting MAPKs and Akt Signaling in RAW264.7 Macrophages. *Biol. Pharm. Bull.* 2018;41(8):1219-27. doi:10.1248/bpb.b18-00037
 103. Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. *Cell Res.* 2005;15(1):11-8. doi:10.1038/sj.cr.7290257
 104. Tay MZ, Poh CM, Rénia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. *Nat. Rev. Immunol.* 2020;20(6):1-12. doi:10.1038/s41577-020-03111-8
 105. Chen G, Wu D, Guo W, et al. Clinical and immunologic features in severe and moderate Coronavirus Disease 2019. *J Clin Invest.* 2020;130(5). doi:10.1172/jci137244
 106. Hojyo S, Uchida M, Tanaka K, et al. How COVID-19 induces cytokine storm with high mortality. *Inflammation and Regeneration* 2020;40(1). doi:10.1186/s41232-020-00146-3
 107. Khezri MR. PI3K/AKT signaling pathway: a possible target for adjuvant therapy in COVID-19. *Hum Cell.* 2021;34(2):700-701. doi:10.1007/s13577-021-00484-5
 108. Miesbach W. Pathological Role of Angiotensin II in Severe COVID-19. *TH Open.* 2020;04(02):e138-e144. doi:10.1055/s-0040-1713678
 109. Marber MS, Rose B, Wang Y. The p38 mitogen-activated protein kinase pathway-A potential target for intervention in infarction, hypertrophy and heart failure. *J Mol Cell Cardiol.* 2011;51(4):485-90. doi:10.1016/j.yjmcc.2010.10.021
 110. Lokhande AS, Devarajan PV. A review on possible mechanistic insights of Nitazoxanide for repurposing in COVID-19. *Eur. J. Pharmacol.* 2021;891:173748. doi:10.1016/j.ejphar.2020.173748
 111. De Rooij J et al. Epac is a Rap1 guanine-nucleotide-exchange factor directly activated by cyclic AMP. *Nature.* 1998;396:474-7. https://doi.org/10.1038/24884
 112. Gray PC, Scott JD, Catterall WA. Regulation of ion channels by cAMP-dependent protein kinase and A-kinase anchoring proteins. *Curr. Opin. Neurobiol.* 1998;8(3):330-4. doi:10.1016/s0959-4388(98)80057-3
 113. Moreno-Fernandez ME, Rueda CM, Velilla PA, Rugeles MT, Chougnat CA. cAMP During HIV Infection: Friend or Foe? *AIDS Res. Hum. Retroviruses.* 2012;28(1):49-53. doi:10.1089/aid.2011.0265
 114. Gerlo S, Verdood P, Kooijman R. Modulation of Cytokine Production by Cyclic Adenosine Monophosphate Analogs in Human Leukocytes. *J Interferon Cytokine Res.* 2010;30(12):883-91. doi:10.1089/jir.2009.0021
 115. Mirza-Aghazadeh-Attari M, Ekrami EM, Aghdas SAM, Mihanfar A, Hallaj S, Yousef B, Safa A, Majidinia M. Targeting PI3K/Akt/mTOR signaling pathway by polyphenols: Implication for cancer therapy. *Life Sci.* 2020;255:117481. doi: 10.1016/j.lfs.2020.11.7481.
 116. George BP, Chandran R, Abrahamse H. Role of phytochemicals in cancer chemoprevention: insights. *Antioxidants (Basel).* 2021;10(9):1455. doi:10.3390/antiox10091455, PMID 34573087.
 117. Maruca A, Catalano R, Bagetta D, Mesiti F, Ambrosio FA, Romeo I et al. The Mediterranean Diet as source of bioactive compounds with multi-targeting anti-cancer profile. *Eur J Med Chem.* 2019;181:111579. doi: 10.1016/j.ejmech.2019.111579, PMID 31398616.
 118. Forni C, Facchiano F, Bartoli M, Pieretti S, Facchiano A, D'Arcangelo D et al. Beneficial role of phytochemicals on oxidative stress and age-related diseases. *BioMed Res Int.* 2019; 2019:8748253. doi: 10.1155/2019/8748253, PMID 31080832.
 119. Vilhelmovali-Ilieva N, Jacquet R, Deffieux D, Pouységu L, Sylla T, Chassaing S, et al. Anti-herpes simplex virus Type 1 activity of specially selected groups of tannins. *Drug Res.* 2019;69(7):374-3. doi:10.1055/a-0640-2557, PMID 30134445.
 120. Cui Q, Du R, Anantpadma M, Schafer A, Hou L, Tian J, et al. Identification of ellagic acid from plant *Rhodiola rosea* L. as an anti-Ebola virus entry inhibitor. *Viruses.* 2018;10(4):152. doi: 10.3390/v10040152, PMID 29584652.
 121. Promsong A, Chuenchitra T, Saipin K, Tewtrakul S, Panichayupakaranant P, Satthakarn S, et al. Ellagic acid inhibits HIV-1 infection *in vitro*: potential role as a novel microbicide. *Oral Dis.* 2018;24(1-2):249-52. doi: 10.1111/odi.12835, PMID 29480632.
 122. Cornélio Favarin D, Martins Teixeira M, Lemos de Andrade E, de Freitas Alves C, Lazo Chica JE, Artério Sorgi C, et al. Anti-inflammatory effects of ellagic acid on acute lung injury induced by acid in mice. *Mediators Inflamm.* 2013; 2013:164202. doi: 10.1155/2013/164202, PMID 23533300.
 123. Pavlova EL, Zografov NN, Simeonova LS. Comparative study on the antioxidant capacities of synthetic influenza inhibitors and ellagic acid in model systems. *Biomed Pharmacother.* 2016;83:755-62. doi: 10.1016/j.biopha.2016.07.046, PMID 27479194.
 124. Sayed AM, Khattab AR, AboulMagd AM, Hassan HM, Rateb ME, Zaid H, et al. Nature as a treasure trove of potential anti-SARSCoV drug leads: a structural/mechanistic rationale. *RSC Adv.* 2020;10(34):19790-802. doi: 10.1039/d0ra04199h, PMID 35685913.

Cite this article: Gupta KK, Narasimhan Y, Ravichandran S, Pemaiah B, Chellappan D, et al. Network Pharmacology-Based Modeling of Phytocompounds in a Traditional *Siddha* Formulation-*Kabasura kudineer* with Special Reference to SARS-CoV-2. *Indian J of Pharmaceutical Education and Research.* 2025;59(2):682-94.